

#### **ORIGINAL ARTICLE**

# Beef cows with larger vulvar width have greater antral follicle count, viable oocytes, and higher circulating AMH

Renata Maculan<sup>1</sup> <sup>(a)</sup>, Gisvani Lopez de Vasconcelos<sup>2</sup> <sup>(a)</sup>, Jesús Alfonso Sánchez Viafara<sup>3,4</sup> <sup>(a)</sup>, Gabriel Miranda Moreira<sup>2</sup> <sup>(b)</sup>, Cintia Vanin<sup>5</sup> <sup>(b)</sup>, Nathalia Alves<sup>5</sup> <sup>(b)</sup>, Marcos Brandão Dias Ferreira<sup>6</sup> <sup>(b)</sup>, José Camisão de Souza<sup>2</sup>\* <sup>(b)</sup>

<sup>1</sup>Instituto Federal do Sul de Minas, Machado, MG, Brasil <sup>2</sup>Departamento de Zootecnia, Universidade Federal de Lavras, Lavras, MG, Brasil <sup>3</sup>Facultad de Ciencias Agrícolas y Veterinarias, Universidad de Santander, Valledupar, Colombia <sup>4</sup>Grupo Investigación y Desarrollo en Sistemas Agropecuarios, Unidad de Investigación Ganadera, Centro de Desarrollo Tecnológico del Cesar, Valledupar, Cesar, Colombia <sup>5</sup>Departamento de Medicina Veterinária, Universidade Federal de Lavras, Lavras, MG, Brasil <sup>6</sup>Empresa de Pesquisa Agropecuária de Minas Gerais, Belo Horizonte, MG, Brasil

**How to cite:** Maculan R, Vasconcelos GL, Viafara JAS, Moreira GM, Vanin C, Alves N, Ferreira MBD, Souza JC. Beef cows with larger vulvar width have greater antral follicle count, viable oocytes, and higher circulating AMH. Anim Reprod. 2025;22(1):e20240077. https://doi.org/10.1590/1984-3143-AR2024-0077

#### Abstract

Owing to the low heritability of reproductive traits, the search for markers and their interrelationship that could indicate reproductively superior individuals is important in the selection process for bovine reproductive efficiency. This study aimed to investigate the possible interrelationships between the antral follicle count (AFC), vulvar-width (VW), anti-Müllerian hormone (AMH) concentrations, fertility in Bos Taurus and Bos Indicus females. Brahman (Bos Taurus-Indicus, n = 126) and Simmental and Angus (Bos Taurus-Taurus, n = 155) cows were classified as having large ( $\geq$ 86 mm) and small (<86 mm) VW. From each group, one blood sample per animal was collected to determine the AMH serum concentrations. The GLIMMIX procedure in SAS® was used to determine whether vulva width (VW) and AMH classes, associated or not with breed, could influence the age at first calving (FCA), calving to first service interval (CFSI), calving interval (CI), number of services per pregnancy (SP), and number of viable oocytes (VO). Antral follicle count (AFC) (36.10  $\pm$  1.90 vs. 22.78  $\pm$  1.64, for large and small VW, respectively), AMH (1.17  $\pm$  0.07 vs. 0.48  $\pm$  0.007 ng/mL), and viable oocytes or VO (18.86  $\pm$  1.76 vs. 10.15  $\pm$  1.49) were greater (*P* < 0.05) in the large VW than in the small VW. Brahman cows had greater AFC ( $36.30 \pm 1.34$  vs.  $22.09 \pm 1.67$ ), VW ( $106.94 \pm 15.83$  vs.  $69.78 \pm 14.11$  mm), and AMH (1.18  $\pm$  0.07 vs. 0.42  $\pm$  0.05 ng/mL) compared to that of taurine cows. In conclusion, VW was an efficient predictor of AFC and AMH concentrations in both genetic groups, but under the conditions of this trial no link could be detected between these variables and the reproductive indices studied.

Keywords: reproductive efficiency, external genitalia, hormonal marker, fertility, viable oocytes.

## Introduction

The selection of bovine females from reproductive indices is limited because of the long generation interval and low heritability of the traits (Silva et al., 2005). Bovine female fertility is measured using reproductive indices such as age at first calving (FCA) and calving interval (CI) (Perotto et al., 2006). However, such indices are subject to the action of non-genetic factors, such as nutrition and management practices (Zink et al., 2011). Therefore, the search for traits that can indirectly indicate reproductively superior animals is essential.

The reproductive structure sizes can affect fertility in bovine females. Ovarian size is a good predictor of follicular reserve and ovarian function (Modina et al., 2014). The external genitalia

\*Corresponding author: jcamisao@ufla.br

Received: June 5, 2024. Accepted: December 18, 2024.

Financial support: RM received funding for this research from Casa Branca Agropastoril, Silvianópolis, Minas Gerais and Fundação de Amparo à Pesquisa de Minas Gerais – FAPEMIG, in partnership with the Universidade Federal de Lavras – UFLA, Lavras, Minas Gerais. Conflicts of interest: The authors have no conflict of interest to declare.

 $\odot$ 

Copyright © The Author(s). This is an Open Access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

(vulva) can predict ovarian follicular reserve and fertility measurements in the field, such as the CI and reproductive efficiency (Mesquita et al., 2016; Maculan et al., 2018). Therefore, it is necessary to investigate the factors that can affect the size of the external genitalia and the possible differences between the genetic groups.

The ovarian follicular reserve is estimated by the antral follicle count (AFC) using ultrasonography. Antral follicle count has a positive association with the results from the use of reproductive biotechnologies (Santos et al., 2016). However, the possible relationships between AFC and reproduction indices remain controversial. Thus, additional studies are necessary to assess the associations of fertility indicator traits (AFC, vulvar measurements, and ovarian volume) with reproductive performance.

Anti-Müllerian hormone (AMH) acts as a modulator of follicular atresia and is an indicator of a greater ovarian follicular reserve (Monniaux et al., 2013).

This study aimed to assess the interrelationships between antral follicle count (AFC), vulvarwidth (VW), anti-Müllerian hormone (AMH) concentrations and reproductive efficiency indices in *Bos Taurus* and *Bos Indicus* females.

## Methods

All procedures and protocols were approved by the Ethics Committee on the Use of Animals (Permanent Commissions/PRP-UFLA) of the Universidade Federal de Lavras, Lavras, Minas Gerais, Brazil (protocol number 063/15).

#### Animals and facilities

The experiment was conducted on the Casa Branca Agropastoril farm (a commercial enterprise) in Careaçu, MG, Brazil. *Bos Indicus* (Brahman breed, n = 124) and *Bos Taurus* (Simmental and Angus breeds, n = 155) were used to identify differences in the genetic group regarding AFC, vulvar width (VW), and AMH concentration. The mean age of zebu females was  $4.8 \pm 2.7$  years (2–9 years) and that of taurine females was  $5.7 \pm 3.2$  years (2–16 years). Zebu cows weighed 317-774 kg (mean of  $531.00 \pm 98.62$  kg), while the taurine cows weighed 258-803 kg (mean of  $533 \pm 95.72$  kg). All cows were on *Brachiaria* sp. Pastures throughout the trial, supplemented with corn silage, *ad libitum* mineral mix, and concentrate at 1.5% of body weight. Reproductive management consisted of natural service and conventional artificial insemination (AI) or fixed-time artificial insemination (FTAI); all cows were cycling and body condition scores (BCSs) ranging from 3 to 8 (1–9, lean to obese; Thomas and Bailey, 2021). The BCS was recorded by a single evaluator. Cows with signs of estrus, those at the end of gestation (30 d to calving), and those in the recent postpartum period (less than 20 d after calving) were excluded from the study. Animals were classified by parity into two groups: N (nulliparous or primiparous, n = 117) and V (multiparous, n = 164).

# AFC and ovarian size

To determine the AFC, the population of antral follicles  $\geq 3$  mm in diameter from both ovaries was assessed using transrectal ultrasonography (Aloka SSD 500; Mure, Japan) with a B-mode linear transducer (5.0 MHz) on a random day of the estrous cycle. *Bos Indicus* cows were classified as high ( $\geq 50$  follicles, n = 37), intermediate (30–49 follicles, n = 70), or low AFC (< 30 follicles, n = 76) (Rodrigues et al., 2015). *Bos Taurus* cows were classified as having high AFC ( $\geq 25$  follicles, n = 39), intermediate AFC (16–24 follicles, n = 16), or low AFC ( $\leq 15$  follicles, n = 44) (Ireland et al., 2007; Burns et al., 2005). Ovarian size (OS) was measured by ultrasonography. All cows were evaluated by the same professional. Ovaries were classified (OSC) according to their longitudinal axes as S (small,  $\leq 2.5$  cm), M (medium, 2.5–3.5 cm), and L (large,  $\geq 3.5$  cm).

#### Morphometry of VW

The morphometry of the external genitalia was measured using a digital caliper (150 mm/0.01 mm Powerfix Nf, Digimess, São Paulo, BR). Vulvar width (VW) was defined as the

distance between the side edges of the vulva and the middle point of the rima length (opening) at an angle of 90° (Figure 1) (Mesquita et al., 2016), based on the frequency distributions of the values obtained for VW (L  $\ge$  86 mm and S < 86 mm).

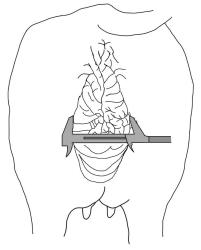


Figure 1. Vulva width measured with a digital caliper.

# **Evaluation of reproductive efficiency**

Reproductive efficiency was evaluated from the reproductive history of females, from the farm records. The variables analyzed were age at first calving (FCA), calving to first service interval (CFSI), calving interval (CI), number of services per pregnancy (SP), and number of viable oocytes (VO) in cows subjected to follicular aspiration protocols.

# **Blood sampling and determination of AMH**

Blood samples were collected by coccygeal venipuncture in 10 mL vacuum tubes (BD VACUTAINER®, São Paulo, BR) on a random day of the estrous cycle. Serum was obtained by centrifugation at 3000 × g for 15 min and were frozen and stored at -20°C for later analyses. Serum AMH concentrations were determined by enzyme-linked immunosorbent assay kit (AnshLabs, Webster, Texas, USA), as previously validated for bovines (Ireland et al., 2008). Three assays were performed with sensitivity of 0.011 ng/mL with intra-assay and inter-assay variabilities from 1.8 to 2.8, and 0.4 to 0.8 respectively. From the respective frequency distributions, the animals were classified as high (>0.81 ng/mL) and low ( $\leq$ 0.81 ng/mL) AMH. The assays were conducted in the IgAc (Instituto Genese de Análises Científicas, São Paulo, BR) laboratory.

# **Statistical analysis**

All statistical analyses were performed using SAS<sup>®</sup> (Cary, NC, USA, 2016). Classes of VW and the AMH concentrations were created based on the respective frequency distributions. General data were tested for normality using the Shapiro–Wilk test according to the UNIVARIATE procedure and, when necessary, transformed to achieve normality. The GLIMMIX procedure was used to assess AFC, FCA, CFSI, CI, SP, and mean number of VO according to BCS, parity, VW class, AMH class, and interactions. Data are presented as mean  $\pm$  standard error of the mean. Differences were considered statistically significant by the Tukey test at *P* < 0.05.

# **Results**

# AFC, reproductive track morphometry, AMH and reproduction

Antral follicle count (AFC) was higher (P < 0.0001) in zebu (36.30 ± 1.34) than in taurine females (22.09 ± 1.67). Variation in the total count ranged from 4 to 119 in zebu females and 1 to 88 in taurine. VW was greater in cows in the high AFC class. AFC was similar between age (P = 0.19) and

parity (*P* = 0.14), regardless of the breed. Ovarian size did not differ between the breeds; however, AFC was higher in the large ovarian class (Table 1).

	OSC						
SMALL	MEDIUM	LARGE	P				
(<2.5 cm)	(2.5-3.5 cm)	(>3.5 cm)					
	AFC						
17.08 ± 13.29 <sup>c</sup>	$29.30 \pm 18.60^{b}$	41.69 ± 27.93 <sup>a</sup>	< 0.05				
( <i>n</i> = 81)	( <i>n</i> = 132)	( <i>n</i> = 62)					
20.20 ± 14.15 <sup>c</sup>	31.16 ± 20.49 <sup>b</sup>	52.34 ± 32.74 <sup>a</sup>	< 0.05				
( <i>n</i> = 121)	( <i>n</i> = 126)	( <i>n</i> = 26)	< 0.05				
VULVAR WIDTH (mm)**							
SMALL	MEDIUM	LARGE					
78.12 ± 2.56 <sup>c</sup> ( <i>n</i> = 82)	88.51 ± 2.02 <sup>b</sup> ( <i>n</i> = 132)	92.70 ± 2.90ª ( <i>n</i> = 64)	<0.001				
82.02 ± 2.12 <sup>c</sup> (n = 121)	87.41 ± 2.08 <sup>b</sup> ( <i>n</i> = 126)	99.16 ± 4.41ª (n = 28)	<0.001				
	$(<2.5 \text{ cm})$ $17.08 \pm 13.29^{\circ}$ $(n = 81)$ $20.20 \pm 14.15^{\circ}$ $(n = 121)$ $SMALL$ $78.12 \pm 2.56^{\circ} (n = 82)$ $82.02 \pm 2.12^{\circ} (n = 121)$	(<2.5 cm)(2.5-3.5 cm)AFC $17.08 \pm 13.29^{c}$ $29.30 \pm 18.60^{b}$ $(n = 81)$ $(n = 132)$ $20.20 \pm 14.15^{c}$ $31.16 \pm 20.49^{b}$ $(n = 121)$ $(n = 126)$ VULVAR WIDTH (mm)**SMALLMEDIUM $78.12 \pm 2.56^{c}$ $88.51 \pm 2.02^{b}$ $82.02 \pm 2.12^{c}$ $87.41 \pm 2.08^{b}$	(<2.5 cm)(2.5-3.5 cm)(>3.5 cm)AFC $17.08 \pm 13.29^{c}$ $29.30 \pm 18.60^{b}$ $41.69 \pm 27.93^{a}$ $(n = 81)$ $(n = 132)$ $(n = 62)$ $20.20 \pm 14.15^{c}$ $31.16 \pm 20.49^{b}$ $52.34 \pm 32.74^{a}$ $(n = 121)$ $(n = 126)$ $(n = 26)$ VULVAR WIDTH (mm)**SMALLMEDIUMLARGE $78.12 \pm 2.56^{c}$ $(n = 82)$ $88.51 \pm 2.02^{b}$ $(n = 132)$ $92.70 \pm 2.90^{a}$ $(n = 64)$				

Table 1. Ovarian size class (OSC)\*, antral follicle count (AFC) and vulvar width (mm).

\*Ovaries were classified (OSC) according to their longitudinal axes as S (small,  $\leq$ 2.5 cm), M (medium, 2.5–3.5 cm), and L (large,  $\geq$ 3.5 cm). \*\* Vulvar width was classified and analyzed over for the right ovary and left ovary separately

In the same row, superscript letters indicate significant differences (P < 0.05). The numbers are the mean values of the least squares ± standard error of the mean.

The VO number was greater (P < 0.05) in the cows of the large (18.86 ± 1.76, n = 38) than in those of the small VW class (10.15 ± 1.49, n = 53).

Reproductive indices (FCA, CFSI, CI, and SP) were similar between AFC classes (P > 0.05).

In zebu, VW was greater (106.94  $\pm$  15.83 mm; *P* < 0.0001) than in taurine cows (69.78  $\pm$  14.11 mm). The VW ranged from 35.15 to 99.24 mm in taurine females and from 67.67 to 143.37 mm in zebu females. Vulvar with (VW) which was smaller (P<0.05) in heifers (81.01  $\pm$  2.17 mm, *n* = 115) than in cows (90.11  $\pm$  1.81 mm, *n* = 165), was greater in larger ovaries (Table 1).

Antral follicle count (AFC) was greater (P < 0.0001) in the large VW class (36.10  $\pm$  1.90 mm) than that of the small class (22.78  $\pm$  1.64 mm). This result was still observed when analyses considered the genetic groups (Table 2).

The AMH concentration in the large VW class (n = 103,  $1.17 \pm 0.07$  ng/mL) was superior (P < 0.0001) to that of the small VW class (n = 119,  $0.49 \pm 0.07$  ng/mL). However, there was an interaction between the genetic groups, such that this result was not maintained in taurine females (Table 2).

	LARGE		SM	SMALL	
	n	AFC	n	AFC	Р
Taurine	11	29.27 ± 5.18ª	144	21.53 ± 1.46 <sup>b</sup>	<0.0001
Zebu	108	$36.80 \pm 2.30^{a}$	17	$33.12 \pm 5.80^{b}$	<0.0001
	n	АМН	n	АМН	
Taurine	11	0.39 ± 0.02 <sup>a</sup>	144	$0.43 \pm 0.05^{a}$	> 0.05
Zebu	108	1.23 ± 0.09 <sup>a</sup>	17	$0.89 \pm 0.02^{b}$	< 0.05

**Table 2.** Vulvar width class (VWC), antral follicle count (AFC) and anti-Müllerian hormone (AMH) concentrations (ng/mL) according to the genetic group

Superscript letters in the same row, superscript letters indicate significant differences (P < 0.05). The numbers are the mean values of the least squares ± standard error of the mean.

The AMH concentration in zebu cows was greater (P < 0.0001) than that in the taurine females (1.18 ± 0.07 ng/mL, n = 112, and 0.42 ± 0.05 ng/mL, n = 110, respectively).

The AMH concentrations did not decrease with age, regardless of the genetic group (P > 0.05), and were similar between parities, regardless of the genetic group (P = 0.84). AMH concentrations

did not differ between BCS in either genetic group (P > 0.05). AMH concentrations in both genetic groups were higher in the large ovary class (Table 3).

**Table 3.** Ovarian size class (OSC) and anti-Müllerian hormone (AMH) concentrations (ng/mL) accordingto the genetic group

	GENETIC GROUP							
	ZEBU				TAURINE			
OSC*	RO	n	LO	n	RO	n	LO	n
Small	1.02 ± 0.02 <sup>c</sup>	31	$1.06 \pm 0.01^{b}$	50	0.29 ± 0.01 <sup>c</sup>	51	$0.36 \pm 0.07^{b}$	71
Medium	$1.25 \pm 0.02^{b}$	63	$1.04 \pm 0.01^{b}$	58	$0.36 \pm 0.09^{b}$	69	$0.53 \pm 0.07^{a}$	68
Large	$1.45 \pm 0.03^{a}$	31	$2.17 \pm 0.02^{a}$	15	$0.68 \pm 0.01^{a}$	33	$0.43 \pm 0.02^{b}$	13

\*Ovarian class in which each ovary (left or right) was included independently of the cow, yielded distinct AMH values. RO: right ovary; LO: left ovary. Superscript letters in the same column indicate significant differences (P < 0.05). The numbers are the mean values of the least squares ± standard error of the mean. Small (<2.5 cm), medium (2.5-3.5 cm), large (>3.5 cm).

There was no difference in reproductive indices between AMH concentration classes (P > 0.05).

The AFC in the zebu cows classified as high AMH was greater (34.54  $\pm$  1.84) than that observed in low AMH zebu cows (23.08  $\pm$  1.74) (*P* < 0.0001). This difference was not observed in taurine cows (Table 4).

**Table 4.** High Anti-Müllerian hormone (AMH) class are related to higher antral follicle counts (AFC) in zebu, but not in taurine females.

		ZEBU			TAURINE	
AMH CLASS <sup>1</sup>	n	AFC	Р	n	AFC	Р
High AMH	78	$40.81 \pm 2.60^{a}$	0.003	52	25.13 ± 2.39 <sup>a</sup>	>0.05
Low AMH	46	$28.65 \pm 3.40^{b}$		99	20.49 ± 1.73 <sup>a</sup>	

<sup>1</sup>High AMH->0.81 ng/mL; Low AMH- $\leq$ 0.81 ng/mL. Superscript letters in the same column indicate significant differences (*P* < 0.05). The numbers are the mean values of the least squares ± standard error of the mean.

# Discussion

Vulvar width in zebu and taurine females was associated with AFC, corroborating previous studies (Mesquita et al., 2016; Maculan et al., 2018). The development of the reproductive tract is influenced by genes (e.g AMH, WNT-A, GATA-4, DHH) which in turn, exert effects on ovarian function including follicular development (Richards, 2001; MacLaughlin et al., 2001). This was evidenced in correlation observed between the ovarian reserve and the development of reproductive tract and external genitalia (Cushman et al., 2013; Honaramooz et al., 2004; Jimenez-Krassel et al., 2009; Maculan et al., 2018; Mesquita et al., 2016).

However, it is not certain whether these positive correlations are due to the action of common genes which stimulate both, folliculogenesis and the development of the reproductive tract or whether it corresponds to specific factors produced by the follicles that exert influence on the reproductive tract development (Cushman et al., 2024).

Additionally, we must consider that, during the reproductive life of the cow, these genes may undergo epigenetic alterations caused by environmental or external factors. Thus, it is possible that cows of similar chronologic age may actually have distinct epigenetic ages. Among these alterations there are distinct ratios between histone deacetylases and methyl transferases abundances, beside DNA methylation (Clarke et al., 2021; Zhu et al., 2023; Klutstein and Gonen, 2023).

Thus, there are cows with characteristics that indicate fertility and others that suggest subfertility due to alterations in the epigenome, causing effects on the follicular cumulus oocyte complex and on the development of the reproductive tract in opposite directions, altering the phenotype match (Cushman et al., 2023; Mossa and Evans, 2023).

Vulvar width (VW) was greater in cows with higher AMH concentrations and AFCs. A direct explanation for this finding it is not clear, however a recently reported AMH effect may help to

elucidate it, which is related to the regulation of the uterine function (Ferdousy et al., 2020). This finding strengthens the already known AMH effects on the quality of bovine oocyte (Vasconcelos et al., 2020; Torres-Rovira et al., 2014; Nguyen et al., 2022) and on the regulation of the ovarian function (Cushman et al., 2002; Gigli et al., 2005; Estienne et al., 2015; Yang et al., 2017; Juengel et al., 2021). Bovine oviducts and uteri have a Type 2-AMH receptor of which no reports on abundance changes throughout the estrous cycle or reproductive senescence have been published yet (Ferdousy et al., 2020). This allows to infer that decreases on AMH circulating concentrations may influence fertility by the regulation of uterine function. Subfertility is related to the depletion of the number of follicles by reproductive aging (Ferdousy et al., 2020).

Lower uterine weight, ovarian weight and endometrial diameter were observed in subfertile ewes and cows (Cushman et al., 2023). This suggests that the size of the reproductive tracts is correlated with fertility since subfertile ewes had lower AMH concentrations.

Moreover, infertile ewes have lower AFC and AMH, and smaller reproductive tracts. However, there are some discrepancies, where there are ewes with ovarian volumes which are in contrast with the fertile/infertile classification, e.g. ewes with larger ovaries and low fertility, and vice versa. Nevertheless, fertile ewes always have superior AMH concentrations (Cushman et al., 2023). In subfertile cows, this correspondence was also observed (Cushman et al., 2013; McNeel et al., 2017; Snider et al., 2022), and lower AFCs were observed in subfertile ewe (Torres-Rovira et al., 2014). This is further supported by the regulation of uterine function by AMH (Ferdousy et al., 2020).

The VW of zebu females was greater than that of taurine females. As mentioned before, AMH is related to fertility and to the development of the reproductive tract (Cushman et al., 2023; Ferdousy et al., 2020), facilitating the understanding of why taurine cows have smaller VW. The overall AMH concentration in zebu cows was higher than in taurine cows. The concentration of AMH is greater in females with intermediate and high AFC. Zebu cows have higher AFC than taurine (Maculan et al., 2018). This may be explained by the greater AMH production occurring in the pre-antral and early antral follicles (Durlinger et al., 2002a). Bos Indicus cows have less primordial follicles compared to Bos Taurus, but the AFC is higher in the former (Silva-Santos et al., 2011). It is believed that AMH participates in the inhibition of the primordial follicle activation mechanism, which triggers its ensuing growth (Durlinger et al., 2002b; Fortune et al., 2011). Thus, it is possible to understand the higher AMH concentrations in Indicus compared to Taurine cows observed in the current report.

The AMH concentration exhibited an association with the number of viable oocytes (VO); however, no association was observed with the reproduction indices in the field and cows classified as having high AMH had higher AFC and with the VO number. It is well established that AMH inhibits the production and follicular sensitivity to follicle-stimulating hormone, thus reducing follicular atresia (Ireland et al., 2009). In other words, AFC increases owing to greater circulating AMH concentrations even in zebu females that do not have a larger number of preantral follicles compared to taurine animals (Silva Santos et al., 2011; Batista et al., 2014). As mentioned before, it may be suggested that a lower atresia rate may lead to a greater number of viable oocytes.

In the present study, ovarian volume was associated with AFC. Ovarian size (OS) may predict the ovarian reserve considering that the hormonal production in the estrous cycle may regulate follicular size (Kwee et al., 2007; Penitente-Filho et al., 2015). Additionally, the gestation period exerts regulatory roles in the OS, which may impact its function and morphology (Chacur et al., 2006).

Antral follicle count is related to various fertility measurements, and the results observed in this study in relation to reproduction are promising, where AFC was associated with a larger OS, VW was associated with high AFC class and in turn AFC was associated with large VW class in both genetic groups.

A direct relationship between ovarian reserve and fertility is still unclear. A number of studies relate higher progesterone concentrations to better oocyte quality and higher follicle counts (Jimenez-Krassel et al., 2009; Martinez et al., 2016; Santa Cruz et al., 2018). However, other reports observed the opposite, where a lower number of follicles linked to higher progesterone concentrations (Lima et al., 2020; Bonato et al., 2022). Moreover, it is important to consider that there is a correlation between the ovarian reserve and external factors, such

as temperature, nutrition and health which induce epigenetic modifications in genes associated to fertility, helping to explain the divergent results related to variations in fertility and their relation with AFC (Cushman et al., 2024).

Antral follicle count was greater in zebu females classified as having high AMH. This association has been clarified in the literature (Rico et al., 2009; Ireland et al., 2010; Batista et al., 2014; Souza et al., 2016; Maculan et al., 2018); however, this relationship was not observed in taurine females in the present study. The AFC and AMH concentrations were highly variable, and the AMH concentration was greater in zebu females. Corroborating these findings, Batista et al. (2014) observed a larger number of follicles and higher AMH concentrations in *Bos Indicus* than in *Bos Taurus* females. Insulin-like growth factor I (IGF-1), in addition to the well-established AMH/AFC link, may help to understand this result, since it is related to higher oocyte quality. Insulin-like growth factor I (IGF-1) concentrations are higher in *Bos Indicus* compared to *Bos Taurus* cows, and this is associated to higher AFC in the former (Alvarez et al., 2000; Satrapa et al., 2013).

#### Conclusion

In conclusion, this study demonstrates that VW is a good predictor of AFC and AMH concentrations in both the genetic groups. Ovarian size, vulvar width, and hormonal traits were positively associated with the number of viable oocytes, antral follicle count and AMH, and can be used in models directed toward the selection of *Bos Taurus Taurus* and *Bos Taurus Indicus* females for superiority in these traits.

#### Acknowledgements

This study was supported and funded by Casa Branca Agropastoril, Silvianópolis, MG, Fundação de Amparo à Pesquisa de Minas Gerais (FAPEMIG), in partnership with the Universidade Federal de Lavras (UFLA), Lavras, Minas Gerais, and the Instituto Genese de Coleções Científicas, São Paulo. We thank the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES) for granting scholarship for doctoral studies.

#### References

- Alvarez P, Spicer LJ, Chase CC Jr, Payton ME, Hamilton TD, Stewart RE, Hammond AC, Olson TA, Wettemann RP. Ovarian and endocrine characteristics during an estrous cycle in Angus, Brahman, and Senepol cows in a subtropical environment. J Anim Sci. 2000;78(5):1291-302. http://doi.org/10.2527/2000.7851291x. PMid:10834585.
- Batista EOS, Macedo GG, Sala RV, Ortolan MD, Sá Filho MF, Del Valle TA, Jesus EF, Lopes RN, Rennó FP, Baruselli PS. Plasma antimullerian hormone as a predictor of ovarian antral follicular population in *Bos indicus* (Nelore) and *Bos taurus* (Holstein) heifers. Reprod Domest Anim. 2014;49(3):448-52. http://doi.org/10.1111/rda.12304. PMid:24689827.
- Bonato DV, Ferreira EB, Gomes DN, Bonato FGC, Droher RG, Morotti F, Seneda MM. Follicular dynamics, luteal characteristics, and progesterone concentrations in synchronized lactating Holstein cows with high and low antral follicle counts. Theriogenology. 2022;179:223-9. http://doi.org/10.1016/j.theriogenology.2021.12.006. PMid:34896744.
- Burns DS, Jimenez-Krassel FJ, Ireland JLH, Knight PG, Ireland JJ. Numbers of antral follicles during follicular waves in cattle: evidence for high variation among animals, very high repeatability in individuals, and an inverse association with serum follicle-stimulating hormone concentrations. Biol Reprod. 2005;73(1):54-62. http://doi.org/10.1095/biolreprod.104.036277. PMid:15744026.
- Chacur MGM, Valentim NC, Martinez AIS, Tostes RA, Kronka SN. Morphometry of ovaries of female zebu Bos Taurus Indicus collected in slaughterhouse. Acta Sci Vet. 2006;34:65-70.
- Clarke S, Caulton A, McRae K, Brauning R, Couldrey C, Dodds K. Beyond the genome: a perspective on the use of DNA methylation profiles as a tool for the livestock industry. Anim Front. 2021;11(6):90-4. http://doi.org/10.1093/af/vfab060. PMid:34934534.
- Cushman RA, Akbarinejad V, Perry GA, Lents CA. Developmental programming of the ovarian reserve in livestock. Anim Reprod Sci. 2024;264:107458. http://doi.org/10.1016/j.anireprosci.2024.107458. PMid:38531261.

- Cushman RA, Miles JR, Rempel LA, McDaneld TG, Kuehn LA, Chitko-McKown CG, Nonneman D, Echternkamp SE. Identification of an ionotropic glutamate receptor AMPA1/GRIA1 polymorphism in crossbred beef cows differing in fertility. J Anim Sci. 2013;91(6):2640-6. http://doi.org/10.2527/jas.2012-5950. PMid:23478821.
- Cushman RA, Wahl CM, Fortune JE. Bovine ovarian cortical pieces grafted to chick embryonic membranes: a model for studies on the activation of primordial follicles. Hum Reprod. 2002;17(1):48-54. http://doi.org/10.1093/humrep/17.1.48. PMid:11756361.
- Cushman RA, Yake HK, Snider AP, Lents CA, Murphy TW, Freking BA. An extreme model of fertility in sheep demonstrates the basis of controversies surrounding antral follicle count and circulating concentrations of anti-Müllerian hormone as predictors of fertility in ruminants. Anim Reprod Sci. 2023;259:107364. http://doi.org/10.1016/j.anireprosci.2023.107364. PMid:37922623.
- Durlinger A, Gruijters MJG, Kramer P, Karels B, Ingraham HA, Nachtigal MW, Uilenbroek JTJ, Grootegoed JA, Themmen APN. Anti-M€ullerian hormone inhibits initiation of primordial follicle growth in the mouse ovary. Endocrinology. 2002b;143(3):1076-84. http://doi.org/10.1210/endo.143.3.8691. PMid:11861535.
- Durlinger A, Visser J, Themmen A. Regulation of ovarian function: the role of anti-Mullerian hormone. Reproduction. 2002a;124(5):601-9. http://doi.org/10.1530/rep.0.1240601. PMid:12416998.
- Estienne A, Pierre A, di Clemente N, Picard JY, Jarrier P, Mansanet C, Monniaux D, Fabre S. Anti-Müllerian hormone regulation by the bone morphogenetic proteins in the sheep ovary: deciphering a direct regulatory pathway. Endocrinology. 2015;156(1):301-13. http://doi.org/10.1210/en.2014-1551. PMid:25322464.
- Ferdousy RN, Kereilwe O, Kadokawa H. Anti-Müllerian hormone receptor type 2 (AMHR2) expression in bovine oviducts and endometria: comparison of AMHR2 mRNA and protein abundance between old Holstein and young and old Wagyu females. Reprod Fertil Dev. 2020;32(8):738-47. http://doi.org/10.1071/RD19121. PMid:32336320.
- Fortune JE, Yang MY, Muruvi W. In vitro and in vivo regulation of follicular formation and activation in cattle. Reprod Fertil Dev. 2011;23(1):15-22. http://doi.org/10.1071/RD10250. PMid:21366976.
- Gigli I, Cushman RA, Wahl CM, Fortune JE. Evidence for a role for anti-Mullerian hormone in the suppression of follicle activation in mouse ovaries and bovine ovarian cortex grafted beneath the chick chorioallantoic membrane. Mol Reprod Dev. 2005;71(4):480-8. http://doi.org/10.1002/mrd.20338. PMid:15895366.
- Honaramooz A, Aravindakshan J, Chandolia RK, Beard AP, Bartlewski PM, Pierson RA, Rawlings NC. Ultrasonographic evaluation of the pre-pubertal development of the reproductive tract in beef heifers. Anim Reprod Sci. 2004;80(1-2):15-29. http://doi.org/10.1016/S0378-4320(03)00136-2. PMid:15036512.
- Ireland JJ, Scheetz D, Jimenez-Krassel F, Folger JK, Smith GW, Mossa F, Evans ACO. Evidence that mammary gland infection/injury during pregnancy in dairy cows may have a negative impact on size of the ovarian reserve in their daughters. Biol Reprod. 2010;83(Suppl 1):277. http://doi.org/10.1093/biolreprod/83.s1.277.
- Ireland JJ, Ward F, Jimenez-Krassel F, Ireland JLH, Smith GW, Lonergan P, Evans ACO. Follicle numbers are highly repeatable within individual animals but are inversely correlated with FSH concentrations and the proportion of good-quality embryos after ovarian stimulation in cattle. Hum Reprod. 2007;22(6):1687-95. http://doi.org/10.1093/humrep/dem071. PMid:17468258.
- Ireland JJ, Zielak-Steciwko AE, Jimenez-Krassel F, Folger J, Bettegowda A, Scheetz D, Walsh S, Mossa F, Knight PG, Smith GW, Lonergan P, Evans AC. Variation in the ovarian reserve is linked to alterations in intrafollicular estradiol production and ovarian biomarkers of follicular differentiation and oocyte quality in cattle. Biol Reprod. 2009;80(5):954-64. http://doi.org/10.1095/biolreprod.108.073791. PMid:19164170.
- Ireland JLH, Scheetz D, Jimenez-Krassel F, Themmen APN, Ward F, Lonergan P, Smith GW, Perez GI, Evans ACO, Ireland JJ. Antral follicle count reliably predicts number of morphologically healthy oocytes and follicles in ovaries of young adult cattle. Biol Reprod. 2008;79(6):1219-25. http://doi.org/10.1095/biolreprod.108.071670. PMid:18768912.
- Jimenez-Krassel F, Folger JK, Ireland JL, Smith GW, Hou X, Davis JS, Lonergan P, Evans AC, Ireland JJ. Evidence that high variation in ovarian reserves of healthy young adults has a negative impact on the corpus luteum and endometrium during estrous cycles in cattle. Biol Reprod. 2009;80(6):1272-81. http://doi.org/10.1095/biolreprod.108.075093. PMid:19211804.

- Juengel JL, Cushman RA, Dupont J, Fabre S, Lea RG, Martin GB, Mossa F, Pitman JL, Price CA, Smith P. The ovarian follicle of ruminants: the path from conceptus to adult. Reprod Fertil Dev. 2021;33(10):621-42. http://doi.org/10.1071/RD21086. PMid:34210385.
- Klutstein M, Gonen N. Epigenetic aging of mammalian gametes. Mol Reprod Dev. 2023;90(12):785-803. http://doi.org/10.1002/mrd.23717. PMid:37997675.
- Kwee J, Elting ME, Schats R, McDonnell J, Lambalk CB. Ovarian volume and antral follicle count for the prediction of low and hyper responders with in vitro fertilization. Reprod Biol Endocrinol. 2007;5(1):9. http://doi.org/10.1186/1477-7827-5-9. PMid:17362511.
- Lima MA, Morotti F, Bayeux BM, de Rezende RG, Botigelli RC, De Bem THC, Fontes PK, Nogueira MFG, Meirelles FV, Baruselli PS, da Silveira JC, Perecin F, Seneda MM. Ovarian follicular dynamics, progesterone concentrations, pregnancy rates and transcriptional patterns in Bos Indicus females with a high or low antral follicle count. Sci Rep. 2020;10(1):19557. http://doi.org/10.1038/s41598-020-76601-5. PMid:33177637.
- MacLaughlin DT, Teixeira J, Donahoe PK. Perspective: reproductive tract development-new discoveries and future directions. Endocrinology. 2001;142(6):2167-72. http://doi.org/10.1210/endo.142.6.8262. PMid:11356658.
- Maculan R, Pinto TLC, Moreira GM, Vasconcelos GL, Sanches JA, Rosa RG, Bonfim RR, Gonçalves TM, Souza JC. Anti-Müllerian hormone (*AMH*), antral follicle count (AFC), external morphometrics and fertility in Tabapuã cows. Anim Reprod Sci. 2018;189:84-92. http://doi.org/10.1016/j.anireprosci.2017.12.011. PMid:29279199.
- Martinez MF, Sanderson N, Quirke LD, Lawrence SB, Juengel JL. Association between antral follicle count and reproductive measures in New Zealand lactating dairy cows maintained in a pasture-based production system. Theriogenology. 2016;85(3):466-75. http://doi.org/10.1016/j.theriogenology.2015.09.026. PMid:26489910.
- McNeel AK, Soares ÉM, Patterson AL, Vallet JL, Wright EC, Larimore EL, Amundson OL, Miles JR, Chase CC Jr, Lents CA, Wood JR, Cupp AS, Perry GA, Cushman RA. Beef heifers with diminished numbers of antral follicles have decreased uterine protein concentrations. Anim Reprod Sci. 2017;179:1-9. http://doi.org/10.1016/j.anireprosci.2017.01.004. PMid:28215453.
- Mesquita NF, Maculan R, Maciel LFS, Alves N, De Carvalho RR, Moreira GM, De Souza JC. Vulvar width and rima length as predictors of the ovarian follicular reserve in bovine females. J Reprod Dev. 2016;62(6):587-90. http://doi.org/10.1262/jrd.2016-059. PMid:27545816.
- Modina SC, Tessaro I, Lodde V, Franciosi F, Corbani D, Luciano AM. Reductions in the number of midsized antral follicles are associated with markers of premature ovarian senescence in dairy cows. Reprod Fertil Dev. 2014;26(2):235-44. http://doi.org/10.1071/RD12295. PMid:23327793.
- Monniaux D, Drouilhet L, Rico C, Estienne A, Jarrier P, Touzé JL, Sapa J, Phocas F, Dupont J, Dalbiès-Tran R, Fabre S. Regulation of anti-Müllerian hormone production in domestic animals. Reprod Fertil Dev. 2013;25(1):1-16. http://doi.org/10.1071/RD12270. PMid:23244824.
- Mossa F, Evans ACO. Review: the ovarian follicular reserve implications for fertility in ruminants. Animal. 2023;17(Suppl 1):100744. http://doi.org/10.1016/j.animal.2023.100744. PMid:37567673.
- Nguyen DK, O'Leary S, Gadalla MA, Roberts B, Alvino H, Tremellen KP, Mol BW. The predictive value of anti-Müllerian hormone for natural conception leading to live birth in subfertile couples. Reprod Biomed Online. 2022;44(3):557-64. http://doi.org/10.1016/j.rbmo.2021.11.018. PMid:35065913.
- Penitente-Filho JM, Jimenez CR, Zolini AM, Carrascal E, Azevedo JL, Silveira CO, Oliveira FA, Torres CA. Influence of corpus luteum and ovarian volume on the number and quality of bovine oocytes. Anim Sci J. 2015;86(2):148-52. http://doi.org/10.1111/asj.12261. PMid:25187422.
- Perotto D, Abrahão JJS, Kroetz IA. Intervalo de partos de fêmeas bovinas Nelore, guzerá x Nelore, Red Angus x Nelore, Marchigiana x Nelore e Simental x Nelore. Rev Bras Zootec. 2006;35(3):733-41. http://doi.org/10.1590/S1516-35982006000300014.
- Richards JS. Perspective: the ovarian follicle-a perspective in 2001. Endocrinology. 2001;142(6):2184-93. http://doi.org/10.1210/endo.142.6.8223. PMid:11356661.
- Rico C, Fabre S, Médigue C, di Clemente N, Clément F, Bontoux M, Touzé JL, Dupont M, Briant E, Rémy B, Beckers JF, Monniaux D. Anti-Müllerian hormone is an endocrine marker of ovarian gonadotropinresponsive follicles and can help to predict superovulatory responses in the cow. Biol Reprod. 2009;80(1):50-9. http://doi.org/10.1095/biolreprod.108.072157. PMid:18784351.

- Rodrigues AS, Oliveira SN, Loiola MVG, Ferraz PA, Chalhoub M, Bittencourt RF, Araujo EAB, Bittencourt TCBSC, Ribeiro AL Fo. Contagem de folículos antrais em fêmeas Nelore submetidas a inseminação artificial em tempo fixo. Cienc Rural. 2015;45(4):711-7. http://doi.org/10.1590/0103-8478cr20140666.
- Santa Cruz R, Cushman RA, Viñoles C. Antral follicular count is a tool that may allow the selection of more precocious Bradford heifers at weaning. Theriogenology. 2018;119:35-42. http://doi.org/10.1016/j.theriogenology.2018.06.010. PMid:29982134.
- Santos GMGD, Silva-Santos KC, Barreiros TRR, Morotti F, Sanches BV, de Moraes FLZ, Blaschi W, Seneda MM. High numbers of antral follicles are positively associated with in vitro embryo production but not the conception rate for FTAI in Nelore cattle. Anim Reprod Sci. 2016;165:17-21. http://doi.org/10.1016/j.anireprosci.2015.11.024. PMid:26711683.
- Satrapa RA, Castilho AS, Razza EM, Pegorer MF, Puelker R, Barros CM. Differential expression of members of the IGF system in OPU-derived oocytes from Nelore (*Bos indicus*) and Holstein (*Bos taurus*) cows. Anim Reprod Sci. 2013;138(3-4):155-8. http://doi.org/10.1016/j.anireprosci.2013.02.023. PMid:23540623.
- Silva JAIIV, Dias LT, Albuquerque LG. Estudo genético da precocidade sexual de Novilhas em um rebanho Nelore. Rev Bras Zootec. 2005;34(5):1568-72. http://doi.org/10.1590/S1516-35982005000500017.
- Silva-Santos KC, Santos GM, Siloto LS, Hertel MF, Andrade ER, Rubin MI, Sturion L, Melo-Sterza FA, Seneda MM. Estimate of the population of preantral follicles in the ovaries of *Bos Taurus* Indicus and *Bos Taurus* Taurus cattle. Theriogenology. 2011;76(6):1051-7. http://doi.org/10.1016/j.theriogenology.2011.05.008. PMid:21722949.
- Snider AP, Crouse MS, Rosasco SL, Epperson KM, Northrop-Albrecht EJ, Rich JJJ, Chase CC Jr, Miles JR, Perry GA, Summers AF, Cushman RA. Greater numbers of antral follicles in the ovary are associated with increased concentrations of glucose in uterine luminal fluid of beef heifers. Anim Reprod Sci. 2022;239:106968. http://doi.org/10.1016/j.anireprosci.2022.106968. PMid:35316712.
- Souza JC, Maculan R, Rosa RG, Smith LF, Casas PS, Faria BRM, Alves N, Gonçalves TM. 130 Antral follicle counts, vulva width, and serum anti-mullerian hormone concentrations in bovine females of the tabapuã breed. Reprod Fertil Dev. 2016;28(2):195. http://doi.org/10.1071/RDv28n2Ab130.
- Thomas J, Bailey E. Body condition score of beef cattle. University of Missoury Extension Bulettin; Missoury; 2021.
- Torres-Rovira L, Gonzalez-Bulnes A, Succu S, Spezzigu A, Manca ME, Leoni GG, Sanna M, Pirino S, Gallus M, Naitana S, Berlinguer F. Predictive value of antral follicle count and anti-Müllerian hormone for follicle and oocyte developmental competence during the early prepubertal period in a sheep model. Reprod Fertil Dev. 2014;26(8):1094-106. http://doi.org/10.1071/RD13190. PMid:24008140.
- Vasconcelos GL, Cunha EV, Maculan R, Sánchez Viafara JA, Barbalho Silva AW, Souza Batista AL, Viana Silva JR, de Souza JC. Effects of vulvar width and antral follicle count on oocyte quality, in vitro embryo production and pregnancy rate in *Bos taurus taurus* and *Bos taurus indicus* cows. Anim Reprod Sci. 2020;217:106357. http://doi.org/10.1016/j.anireprosci.2020.106357. PMid:32408964.
- Yang MY, Cushman RA, Fortune JE. Anti-Müllerian hormone inhibits activation and growth of bovine ovarian follicles in vitro and is localized to growing follicles. Mol Hum Reprod. 2017;23(5):282-91. http://doi.org/10.1093/molehr/gax010. PMid:28333275.
- Zhu Z, Xu W, Liu L. Ovarian aging: mechanisms and intervention strategies. Med Rev. 2023;2(6):590-610. http://doi.org/10.1515/mr-2022-0031. PMid:37724254.
- Zink V, Štípková M, Lassen J. Genetic parameters for female fertility, locomotion, body condition score, and linear type traits in Czech holstein cattle. J Dairy Sci. 2011;94(10):5176-82. http://doi.org/10.3168/jds.2010-3644. PMid:21943767.

#### Author contributions

RM, GLV, JASV, GMM, CV, NA, MBDF and JCS: Conceptualization, Investigation, Project administration, Writing – original draft; CV and NA: Data curation; RM, GLV, JASV, GMM, CV, NA, MBDF and JCS: Investigation; GLV, JASV, GMM, CV and NA: Methodology; RM and MBDF: Project administration; GLV, JASV, GMM, CV, NA and MBDF: Resources; MBDF, JASV and GMM: Supervision; GMM, CV, NA, MBDF and JCS: Validation; RM, JASV and JCS: Writing – original draft.