



Mechanisms regulating follicle selection in ruminants: lessons learned from multiple ovulation models

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Abstract

Selection of a single dominant follicle from a cohort of growing follicles is a unique biological process, a key step in female reproductive function in monovular species, and lies at the core of reproductive technologies in cattle. Follicle growth and the number of follicles that ovulate are regulated by precise endocrine, paracrine, and autocrine mechanisms. Most of our current understanding about follicle selection focuses on the role of FSH, LH, and the IGF family in follicle growth and selection of the dominant follicle. However, more recently the role of members of the TGF- β family has been highlighted, particularly in high fecundity genotypes in sheep. Intercellular signaling between the oocyte and granulosa cells (GC) regulates proliferation and differentiation due to actions of bone morphogenetic protein 15 (BMP15) and growth and differentiation factor 9 (GDF9) within the follicle. Mutations that either knockout or reduce the activity of BMP15 or GDF9 have been found to increase ovulation rate in heterozygotes and generally cause severe follicle abnormalities in homozygotes. A mutation in the intracellular kinase domain of the BMPRI1B receptor (Booroola fecundity gene) increases ovulation rate in heterozygotes with further increases in ovulation in homozygotes. The physiological mechanisms linking these mutations to increased ovulation rates are still not well defined. A recently identified high fecundity bovine genotype, Trio, causes increased expression of SMAD6, an intracellular inhibitor of the BMP15/GDF9 signalling pathways. This bovine model has provided insights into the mechanisms associated with selection of multiple dominant follicles and multiple ovulations in carriers of fecundity alleles. The present review focuses on the mechanisms involved in follicle selection in ruminants with a special emphasis on the contribution made by multiple ovulation models in both cattle and sheep. The evaluation of multiple ovulation models in ruminants has allowed us to construct a new physiological model that relates changes in the BMP15/GDF9 signalling pathways to the physiological changes that result in selection of multiple dominant follicles. This model is characterized by acquisition of dominance at a smaller follicle size but at a similar time in the follicular wave with multiple follicles acquiring dominance in a hierarchical sequence, delaying FSH suppression and, thus allowing additional follicles to continue to grow and acquire dominance.

Keywords: follicle selection, high fecundity, ruminants.

Introduction

Selection of a single dominant follicle from a cohort of growing follicles is a unique biological process and a key step in female reproductive function. Disturbances in this process can lead to anovulation and infertility or, alternatively, multiple ovulations and multiple births. The improvement in our understanding of follicle development and selection has fueled the development of synchronization protocols for fixed time artificial insemination as well as the development of other reproductive biotechnologies such as superovulation and embryo transfer. As a result, these advances have highlighted the importance of optimal follicle growth and selection as a critical step towards achieving reproductive efficiency in livestock species in order to feed a growing world population.

Alterations in follicle selection can lead to the occurrence of codominant follicles and multiple ovulations which are the basis for dizygotic twinning in cattle and sheep. A particularly useful approach, suitable for study of many biological processes, is the examination of abnormal phenotypes. In this regard the occurrence of multiple ovulations in otherwise monovular species provides a physiological model in which the follicle selection mechanism has been altered in such a way to allow multiple follicles to be selected and ovulate. The available multiple ovulation models in sheep (Juengel *et al.*, 2013) and, more recently, in cattle (Kamalludin *et al.*, 2018) have contributed to the identification of members of the transforming growth factor- β (TGF- β) superfamily as key regulators of ovulation rate and prolificacy unveiling a new set of genes, pathways, and autocrine mechanisms with critical roles in the ovarian physiology of ruminants.

The term follicle selection has been widely used in the literature, however there is a lack of consensus on its scope and implications for reproductive function. Selection has been used in relation to terminal growth of antral follicles, however the loss of preantral follicles by atresia during the early stages of folliculogenesis could arguably be included as part of the selection mechanism. Ginther *et al.* (2001) provided a precise definition for the term in relation to monovular species: is the process wherein one follicle develops from a wave of growing follicles and becomes the only follicle with ovulatory capacity. However, this definition does not account for the possibility of

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multiple follicles being selected. Conversely, the definition of the word selection is: a collection of things chosen from a group of similar things. Therefore, in order to determine whether follicle selection is occurring, two related events need to be observed: 1) a cohort of growing follicles, and 2) the ability of one (or multiple) follicles to continue to grow and acquire the capacity to ovulate while the remaining follicles do not. In the present review we will use the latter definition which provides the opportunity for more than one follicle to be selected, and thus the occurrence of multiple ovulations can be studied in relation to the follicle selection mechanism.

The purpose of this review is to briefly explore the follicular, endocrine and paracrine factors associated with selection of the dominant follicle and more importantly to explore the contribution of multiple ovulation models in ruminants for understanding the follicle selection mechanism. After introducing the genetic models that have been found to increase ovulation rate in ruminants, the review will explore the potential physiology that underlies the increased ovulation rate in both ovine and bovine models.

Selection of a single dominant follicle

Follicle development is an essential aspect of female reproduction with intricate mechanisms driving all stages of this process. Initial follicle development involves gradual activation of primordial follicles, which occurs throughout the reproductive lifespan of the female. The complex mechanisms involved in primordial follicle activation are slowly becoming clearer with a key role for KIT ligand from the granulosa cells (GC) activating PI3K/Akt/mTOR pathways in the oocyte and eventually blocking the transcriptional machinery that tonically inhibits primordial follicle activation, such as Foxo3a and p27 (Zhang and Liu, 2015; Bertoldo *et al.*, 2018; Kallen *et al.*, 2018). Subsequent growth of the primary, secondary, and early antral follicles involves FSH and a series of paracrine factors, such as C-type natriuretic peptide, that stimulate follicle growth allowing the growing follicles to overcome inhibitory pathways, such as the Hippo pathway, ultimately allowing follicles to enter the antral stages of follicle development (Hsueh *et al.*, 2015). The final development stages of antral follicle growth occurs in a wave-like pattern in ruminants, a model first proposed by Rajakoski (1960). The introduction of ultrasonography and the resulting ability to image the bovine ovary allowed for the concept of follicular waves to be revisited and investigated (Pierson and Ginther, 1987; Sirois and Fortune, 1988). Ultrasonography has now provided substantial information on the dynamics of follicle development and has provided convincing data supporting the follicular wave concept. Moreover, the ability to monitor follicle growth patterns has provided the ability to study follicle selection from a dynamic perspective and allow for the evaluation of temporal associations between follicle events and endocrine changes.

A follicular wave is defined as the synchronous growth of a group of small antral follicles, from which a single follicle is selected becoming the dominant follicle, whereas the remainder of the follicles (subordinate follicles) undergo regression (Ginther *et al.*, 1989; Adams, 1994). The follicular wave pattern in ruminants is characterized by the development of typically 2 or 3 follicular waves in cattle (Knopf *et al.*, 1989), and 3 to 6 waves in sheep (Ginther *et al.*, 1995), during each cycle. Traditionally the emergence of the follicular wave has been defined, in cattle, as the day or examination at which the retrospectively identified dominant follicle is first detected at a diameter of 4-5 mm (Ginther *et al.*, 1989).

The pivotal event leading to the occurrence of a single ovulation in monovular species, such as cattle, is referred to as follicle selection. The morphological visualization of the follicle selection process has been termed, diameter deviation, and consists of the continuous growth of the future dominant follicle while the subordinate follicles slow their growth rate or completely cease to grow (Ginther *et al.*, 1996, 1997a, 2001). In cattle, diameter deviation occurs, on average, between 2 and 3 days after emergence of the follicular wave, and, although there is individual variability, this range appears to be very representative as it has been observed in multiple studies using both *Bos taurus* and *Bos indicus* breeds (Ginther *et al.*, 1996; Sartori *et al.*, 2001; Gimenes *et al.*, 2008). The future dominant follicle is on average 8 to 9 mm at deviation while the largest subordinate follicle is 7 to 8 mm in *Bos taurus* (Ginther *et al.*, 1996, 1997a). Conversely, in *Bos indicus* the future dominant follicle and largest subordinate follicle at deviation onset are 5.9 to 7 mm and 5.4 to 5.9 mm, respectively (Sartorelli *et al.*, 2005; Gimenes *et al.*, 2008; Sartori *et al.*, 2016). These data support the idea that the future dominant follicle generally has a size advantage over the largest subordinate follicle. In this regard, a study in *Bos taurus* found that in 76% of 33 waves the future dominant follicle was larger at deviation than the largest subordinate follicle, in 21% they had the same diameter, while in only 3% of the waves the future dominant follicle was smaller (Ginther *et al.*, 1997a). Interestingly, despite diameter deviation occurring at similar times after wave emergence in *Bos taurus* and *Bos indicus*, the size of both the future dominant follicle and the largest subordinate are significantly smaller in *Bos indicus*. The precise mechanism that causes *Bos indicus* to have a smaller follicle size at the time of deviation has not been fully elucidated.

Activation of diameter deviation occurs rapidly (<8 h), as shown by the inhibition of follicle growth less than 8 h after FSH suppression and the growth cessation of the largest subordinate follicle within 8 h after the future dominant follicle reaches 8.5 mm in diameter (Ginther *et al.*, 1999). As the follicular wave develops in response to the FSH surge, the developing follicles themselves secrete FSH suppressors, mainly inhibin, that causes the circulating FSH to decline as the wave progresses (Gibbons *et al.*, 1999b). The role of FSH in the selection process can be summarized through what



has been termed the two-way functional coupling hypothesis between the follicles and FSH. This hypothesis states that during the common growth phase all follicles within the wave contribute (i.e. inhibin) to suppress FSH while depending on FSH for continued growth. Once diameter deviation occurs, the future dominant follicle assumes the coupling role by providing the final suppression of FSH, causing it to reach its nadir while acquiring the ability to survive in the face of basal FSH concentrations (Ginther *et al.*, 1996, 1997a, 2000b; Ginther, 2000). Evidence for this hypothesis has been provided by the following results: 1) deviation occurs in association with FSH concentration reaching its nadir (Ginther *et al.*, 1996, 1997a; Ginther, 2000); 2) initially all the follicles within the wave contribute to the suppression of FSH as seen by the increase in FSH when a portion or all the follicles are ablated (Gibbons *et al.*, 1997); 3) depression of FSH concentrations during the common growth phase by treatment with estradiol (E2) inhibits the growth of all follicles in the wave (Ginther *et al.*, 2000b); 4) administration of exogenous FSH allows for continuous growth of follicles and rescue of those destined to become subordinate allowing for multiple ovulations (Adams *et al.*, 1993); 5) administration of an inhibin antiserum early in the follicular wave results in development of more than one dominant follicle (Kaneko *et al.*, 1993; Takedomi *et al.*, 1997); 6) removal of the dominant follicle after deviation is followed by an increase in FSH within 1 h and subsequent increase in the diameter of the largest subordinate follicle occurring 3 h after the increase in FSH (Ginther *et al.*, 2016); 7) reduction of FSH concentrations at the time of deviation, by either treatment with a steroid-free fraction of follicular fluid or E2, led to a reduction in the growth of the dominant follicle but not the subordinate follicles (Bergfelt *et al.*, 2000; Ginther *et al.*, 2000b); and 8) treatment with E2 antiserum leads to an increase in circulating FSH and a delay in deviation (Beg *et al.*, 2003).

Circulating luteinizing hormone (LH) has also been implicated in follicle selection. Perhaps the most compelling piece of evidence for the role of LH in follicle selection is the lack of continuous growth of the dominant follicle beyond deviation in the absence of LH pulses (Fike *et al.*, 1997; Haughian *et al.*, 2013). For example, treatment with acyline (GnRH antagonist) during the first follicular wave reduced circulating LH concentrations and prevented diameter deviation and growth of the largest follicle past 9 mm. Interestingly, the reduction of circulating LH did not affect follicle growth up to the onset of diameter deviation (Haughian *et al.*, 2013).

Circulating E2 concentrations begin to increase at the time of deviation as a result of increased E2 production by GC of the future dominant follicle (Ginther *et al.*, 1997b; Kulick *et al.*, 1999). As previously discussed, the increase in E2 contributes to the final suppression of FSH, such that ablation of the dominant follicle at deviation (~8.5 mm) causes an increase in FSH and, if exogenous E2 is administered, the increase in FSH is delayed for a period of time

associated with the increase in E2 (Ginther *et al.*, 2000a).

Follicle selection and acquisition of dominance not only involves morphological and endocrine changes but also profound changes in follicular fluid, GC and theca cells (TC). Among the intra-follicular components, E2 concentration has been one of the best characterized changes associated with diameter deviation and acquisition of dominance. Intrafollicular E2 has been shown to increase in the future dominant follicle at or immediately after deviation, and this is associated with increased mRNA expression for CYP19A1 (aromatase) in GC (Beg *et al.*, 2000; Luo *et al.*, 2011).

Induction of LH receptors (LHCGR) in GC has been proposed as one of the initial signatures of the dominant follicle phenotype (Beg *et al.*, 2001; Luo *et al.*, 2011). The induction of LHCGR in GC appears to be stimulated, at least in part, by FSH and is mediated by increases in cAMP (Luo *et al.*, 2003; Nogueira *et al.*, 2007). First detection of a difference in LHCGR between the future dominant follicle and the largest subordinate was at 8 to 8.4 mm, immediately prior to diameter deviation at ~8.5 mm (Beg *et al.*, 2001). More interestingly, it appears that LH pulses are required for induction of LHCGR in GC, as demonstrated by the lack of LHCGR after treatment with acyline 24 h prior to expected deviation (Luo *et al.*, 2011). Follicles that have acquired dominance are the only follicles that ovulate after an LH surge, termed ovulatory capacity, and this capacity is preceded by the acquisition of LH receptors in GC (Sartori *et al.*, 2001). Acquisition of ovulatory capacity occurs just after follicle selection in *Bos taurus* and *Bos indicus*, even though the size of the selected dominant follicle is quite different (Sartori *et al.*, 2001; Gimenes *et al.*, 2008; Simões *et al.*, 2012). Two other factors that are involved in acquisition of follicle dominance are free IGF1, which decreases in the subordinate follicle but remains elevated in the dominant follicle (Beg *et al.*, 2000, 2001) due to breakdown of IGF binding proteins (IGFBP) by the IGFBP protease, PAPPA (Rivera and Fortune, 2003) and FGF10 or FGF18 which increase in the subordinate follicles (Gasperin *et al.*, 2012; Portela *et al.*, 2015).

Multiple ovulation models in ruminants

Genetic models of multiple ovulation

The Booroola Merino ewe was the first high prolificacy line described. It originated from a flock of the Commonwealth Scientific and Industrial Research Organization (CSIRO) using triplet, quadruplet, and quintuplet born ewes and a quintuplet born ram obtained from a commercial operation, Booroola, owned by the Sears Brothers in Cooma, NSW, Australia in 1958 (Bindon, 1984). Since then, at least 19 different mutations affecting ovulation rate related to 6 different genes (Table 1) have been described in sheep and more recently a new mutation has been identified in cattle (Kirkpatrick and Morris, 2015).



Table 1. High fecundity genotypes in sheep showing affected gene, allele, line and breed, reported ovulation rate and percentage increase over wild type allele in heterozygous and homozygous carriers and proposed functional modifications in the proteins.

Gene	Mutated allele	Name/Breed	Number of ovulations (% increase relative to controls)		Ref.
			Heterozygous	Homozygous	
BMPR1B	FecB ^B	Booroola	2.8 (+85%)	4.6-9.7 (+204% to 439%)	Davis <i>et al.</i> , 1982; Mulsant <i>et al.</i> , 2001; Wilson <i>et al.</i> , 2001; McNatty <i>et al.</i> , 2017
	-	Mehraban	1.3† (+25%)	NR	Abdoli <i>et al.</i> , 2013
BMP15	FecX ^I	Inverdale (Romney)	2.5-3.2 (+35% to 64%)	POF (primary stage)	Braw-Tal <i>et al.</i> , 1993; Shackell <i>et al.</i> , 1993; Galloway <i>et al.</i> , 2000; Davis <i>et al.</i> , 2001a
	FecX ^H	Hanna (Romney)	2.6-3.2 (+46% to 72%)	POF (primary stage)	Galloway <i>et al.</i> , 2000; Davis <i>et al.</i> , 2001a; Hanrahan <i>et al.</i> , 2004
	FecX ^B	Belclare (Belclare)	3.3 (+70%)	POF	Hanrahan <i>et al.</i> , 2004
	FecX ^G	Galway (Belclare, Cambridge)	2.7-3.1 (+37 to 42%)	POF	Hanrahan <i>et al.</i> , 2004; McNatty <i>et al.</i> , 2004
	FecX ^L	Lac X-mutated (Lacaune)	3.3-7.2 (+69% to +269%)	POF (primary stage)	Bodin <i>et al.</i> , 2007; Drouilhet <i>et al.</i> , 2009
	FecX ^R	Raza Aragonesa	2.0 (+46%)	POF	Martinez-Royo <i>et al.</i> , 2008; Lahoz <i>et al.</i> , 2011
	FecX ^{Gr}	Grivette	2.9 (+16%)*	4.6 (+81%)	Demars <i>et al.</i> , 2013
	FecX ^O	Olkuska	2.0 (+32%)	3.3 (+142%)	Demars <i>et al.</i> , 2013
	FecX ^{Bar}	Tunisian Barbarine	1.8 (+64%)	POF	Lassoued <i>et al.</i> , 2017
GDF9	FecG ^H	High Fertility (Belclare, Cambridge)	4.3 (+88%)	POF	Hanrahan <i>et al.</i> , 2004
	FecG ^T	Thoka (Icelandic)	+32% (lambing rate)	POF (primary/secondary)	Nicol <i>et al.</i> , 2009; Juengel <i>et al.</i> , 2013
	FecG ^E	Embrapa (Santa Ines)	1.3 (+10%)*	2.2 (+82%)	McNatty <i>et al.</i> , 2004; Silva <i>et al.</i> , 2011
	FecG ^V	Vacaria (Ile de France)	2.4-2.5 (+94%)	POF (small antral follicles abnormal)	Souza <i>et al.</i> , 2014
	FecG ^F	Finnsheep	2.48 (+6%)	2.98 (+28%)	Vage <i>et al.</i> , 2013; Mullen and Hanrahan, 2014
B4GALNT2 [†]	FecL ^L	Lacaune	3.1 (+114%)	4.6 (+214%)	Drouilhet <i>et al.</i> , 2009
Unknown	FecX ^{2W}	Woodlands (Coopworth)	2.7 (+25%)	NR - fertile	Davis <i>et al.</i> , 2001b
Unknown	FecW	Wishart (Romney)	+0.8-1.0 ovulations	NR- fertile	Davis <i>et al.</i> , 2006
Unknown	FecD	Davisdale (Border Leicester X Romney)	+0.4-0.8 ova	NR - fertile	Juengel <i>et al.</i> , 2011

*Non-significant increases from wild-type controls. POF = Primary ovarian failure. NR = not reported.

*Ovine models - Role of TGF- β family members*

The evaluation of the aforementioned high ovulation rate phenotypes in sheep has led to the identification of specific genes and pathways with previously unknown effects on ovulation rate. Single gene mutations such as Booroola/FecB, Inverdale/FecX^L, Hannah/FecX^H and FecG^H led to the discovery of the role of TGF- β signaling in folliculogenesis and ovulation rate. The lines listed above resulted in the identification of two oocyte-secreted factors (OSF), namely bone morphogenetic protein 15 (BMP15), and growth and differentiation factor-9 (GDF9), as key regulators of GC function (Juengel *et al.*, 2013). The original Booroola mutation had a non-conservative substitution in the intracellular kinase domain of the BMPR1B receptor (Mulsant *et al.*, 2001; Souza *et al.*, 2001; Wilson *et al.*, 2001), affecting subsequent signaling pathways. Other mutations identified in sheep result in absent, non-functional, or modified forms of BMP15 or GDF9 (Galloway *et al.*, 2000; Hanrahan *et al.*, 2004).

BMP15 and GDF9 are primarily OSF (Juengel *et al.*, 2002), translated as larger precursor proteins, with a pro-region required for proper folding. Upon cleavage by furin-like proteases the mature dimeric form of the protein is produced (McNatty *et al.*, 2004; Weiss and Attisano, 2013). Interestingly, GDF9 and BMP15 have been shown *in vitro* to form homodimers and heterodimers, however, unlike most TGF- β members these are not covalently linked by disulfide bonds (Liao *et al.*, 2003). As TGF- β members, both BMP15 and GDF9 signal through an assembly of type-I and type-II receptors into heterotetrameric complex receptors (Fig. 1). The type II receptor BMPRII is common to both (Moore *et al.*, 2003), however, GDF9 and BMP15 differ in the type I receptor utilized, with GDF9 signaling through ALK5/TGF β RI or ACVR1B/ALK4 (Li *et al.*, 2011; Peng *et al.*, 2013) and BMP15 signaling through ALK6/BMPRIB (Moore *et al.*, 2003). Upon BMP15 binding to its receptor, phosphorylation of the type-I receptor (ALK6/BMPRIB) occurs by means of the kinase domain of the type II receptor, which is constitutively active, ultimately leading to phosphorylation of receptor-regulated SMAD1/5/8 (R-SMAD) and formation of heteromeric complexes of activated R-SMADs with SMAD4 which accumulate in the nucleus and affect gene expression in a cell-type dependent manner (ten Dijke and Hill, 2004; Weiss and Attisano, 2013). Conversely, GDF9 receptor type I (TGF β RI/ALK5) is phosphorylated upon ligand binding and results in the activation of a different set of R-SMADs (SMAD2/3) with subsequent formation of heteromeric complexes with SMAD4 (Gilchrist *et al.*, 2006). Two inhibitory SMAD proteins have been described, SMAD6 and SMAD7, which act as negative modulators of BMP15 and GDF9 signalling (Li, 2015). Knockdown of SMAD7 significantly enhanced expression of GDF9-stimulated genes indicating that SMAD7 appears to preferentially inhibit the SMAD2/3 pathway. Conversely, SMAD6 preferentially inhibits BMP signaling by several proposed mechanisms: 1)

acting as a SMAD4 decoy thus reducing the formation of SMAD1-SMAD4 heteromers (Hata *et al.*, 1998); 2) interaction with type I receptors (e.g BMPR1B) preventing phosphorylation of SMAD1/5/8 (Imamura *et al.*, 1997; Goto *et al.*, 2007); 3) acting as an adaptor protein for Smad ubiquitin regulatory factor 1 (Smurf1) which leads to ubiquitination and degradation of type I receptors and R-SMADs (Murakami *et al.*, 2003). The high fecundity alleles that act through the GDF9/BMP15 pathways are shown in Fig. 1 (n = 16).

Recently, GDF9 and BMP15 have been shown to act synergistically to regulate GC function, through activation of their receptor complexes as homodimers or heterodimers (Liao *et al.*, 2003; McNatty *et al.*, 2004). A series of studies have shown that addition of both, recombinant BMP15 (murine, human or ovine) and GDF9 (murine, human or ovine) to murine or rat GC produced a greater effect in GC proliferation than either one alone (Reader *et al.*, 2011, 2016; Mottershead *et al.*, 2012). Moreover, this effect appears to be mediated by SMAD2/3 activation rather than SAMD1/5/8. Another study also evaluated the effect of recombinant non-purified murine and ovine GDF9 and BMP15 on rat GC (McNatty *et al.*, 2005a). Thymidine incorporation indicated that murine GDF9 was the only factor that stimulated proliferation on its own, while ovine GDF9 or BMP15 alone had no effect. However, the combination of murine GDF9 with ovine BMP15 or ovine BMP15 with ovine GDF9 had a dose-dependent synergistic effect on thymidine incorporation, much greater than murine GDF9 alone. In addition, these authors demonstrated that either factor alone had no effect on progesterone (P4) production, but mGDF9+oBMP15 or oGDF9+oBMP15 significantly reduced P4 production in a dose-dependent manner.

The individual and cooperative role of BMP15 and GDF9 has also been investigated in ruminant GC (McNatty *et al.*, 2005b). Granulosa cell proliferation was stimulated by oBMP15 and oBMP15+oGDF9 to similar levels in bovine GC, while oGDF9 had no effect and intriguingly mGDF9 inhibited bovine GC proliferation. In ovine GC, addition of the aforementioned factors yielded similar results as those observed in bovine GC; however, the addition of both GDF9 and BMP15 regardless of species of origin resulted in a significantly greater effect on proliferation than with either factor alone. Progesterone production by bovine GC was inhibited by oBMP15 and oGDF9 alone at the highest dose, while combination of both greatly inhibited P4 secretion. In sheep GC, oBMP15 was without effect on P4 secretion, however oGDF9 inhibited while mGDF9 stimulated secretion, and addition of oBMP15 to GDF9 of either species did not modify the effects observed with GDF9 alone (McNatty *et al.*, 2005b).

The results previously described emphasize that: 1) both GDF9 and BMP15 regulate GC proliferation and differentiation measured through P4 production *in vitro*; 2) the effect of each factor depends on the species of origin for BMP15 and GDF9, as well as the species of origin of the GC; and 3) a cooperative effect between BMP15 and GDF9 has been observed in

all species evaluated, albeit differences appear to exist among species regarding which particular GC function is affected by the cooperation. In addition, recent results indicate that BMP15 and GDF9 proteins can occur as dimers, monomers, and multimers and that components of the immature protein such as the pro-region may have important roles in the biological effects of these OSF (Reader *et al.*, 2011; Heath *et al.*, 2017). Further, it has been suggested that the dimerization of BMP15 and

GDF9 may be occurring at the time of receptor binding with minimal dimerization prior to secretion from GC or within the follicular fluid (Heath *et al.*, 2017). It seems clear that further research is needed in this important area in order to establish the precise roles of BMP15 and GDF9 and their dimers in regulation of GC function, particularly with respect to species differences and the possibility that these differences may underlie differences in the number of follicles selected.

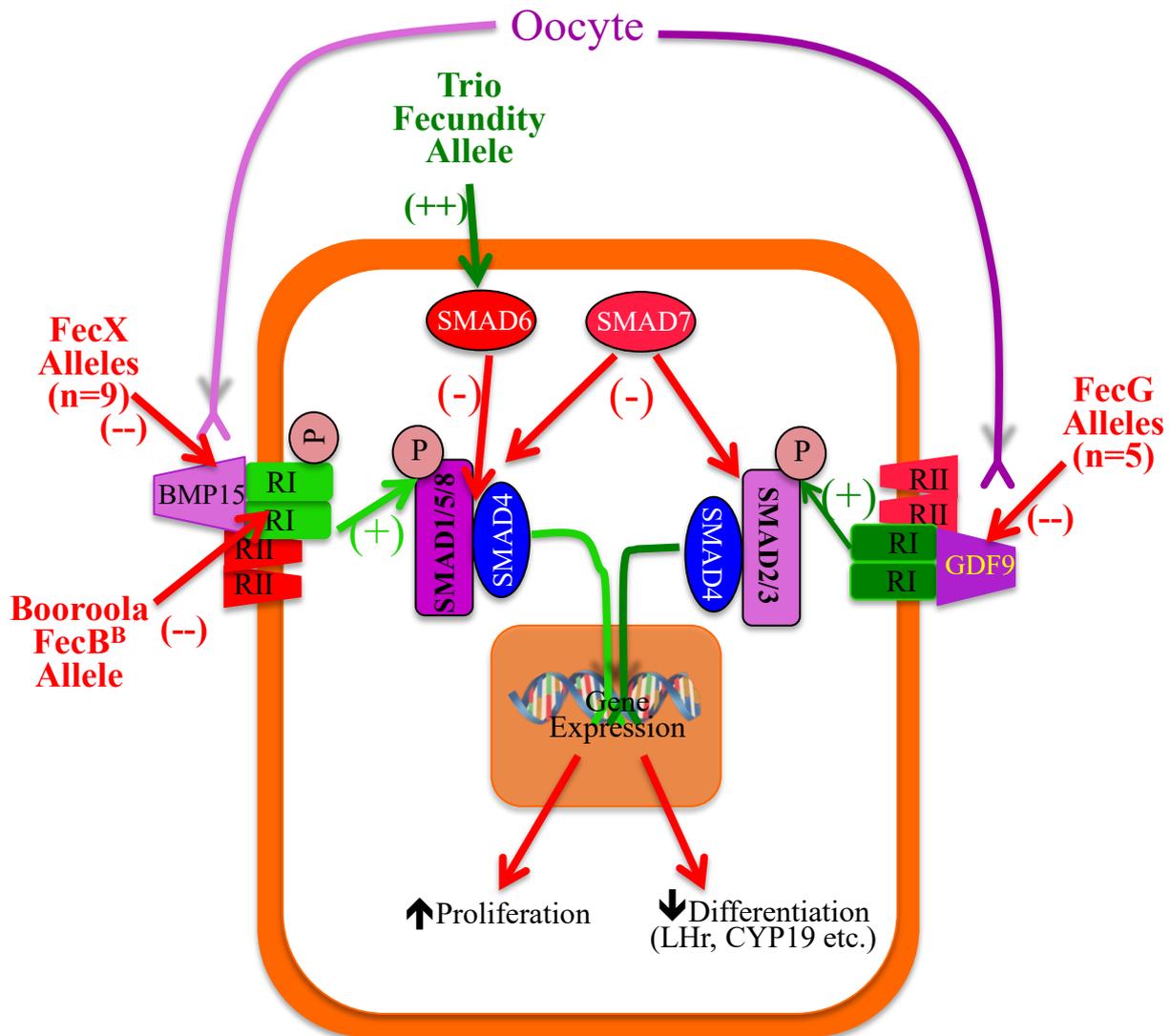


Figure 1. Our current working model of mechanisms in granulosa cells (GC) that lead to reduced follicle growth rate and earlier follicular dominance in carriers of some (n = 16) high fecundity alleles that have been identified in ruminants. The ovine FecX alleles (n = 9) alter the activity or knockout the BMP15 protein; whereas, ovine FecG alleles (n = 5) alter the activity or knockout the GDF9 protein. Both of these proteins are members of the TGF- β family that are secreted by the oocyte as homodimers or heterodimers and regulate independently or, more likely, cooperatively the proliferation and function of the GC. The ovine Booroola FecB allele causes an alteration in the kinase region of the BMPRII receptor and thus reduces the activity of the BMP15 pathway. Carriers of the novel bovine Trio allele have a mutation in a regulatory region for SMAD6 causing a dramatic increase in expression of SMAD6 in GC with subsequent inhibition of BMP-15 action by inhibiting Smad-1/5/8 and thus reduces proliferation and allows differentiation.

Bovine models

Multiple ovulation models in cattle can be divided into three main types, namely: the USDA-MARC twinner population (Echternkamp *et al.*, 2004),

the high producing lactating dairy cow (Lopez *et al.*, 2005), and the recently identified Trio high fecundity allele (Kirkpatrick and Morris, 2015). Genomic contributions to twinning rate in cattle have been studied by QTL mapping within paternal half-sib



families and genome-wide association analyses (GWAS). QTL mapping analyses typically allowed identification of genomic regions potentially associated with twinning rate with broad confidence intervals, making identification of causative genes and polymorphisms extremely challenging. GWAS, by exploiting association across a population rather than within family, potentially narrows localization of underlying genes, though validation is critical. Multiple positional candidate gene regions have been identified by QTL and GWAS analyses (Table 2), though only a few have been replicated.

One of the most replicated regions is the bovine chromosome 5 segment containing IGF1. QTL mapping work in both the Norwegian dairy cattle population and the US Holstein population has provided strong support for a contribution of this genomic region to variation in twinning rate. In a follow-up study, polymorphisms in the IGF1 gene were identified and tested for association in two samples of US Holstein sires representing different timeframes. IGF1 polymorphisms were identified with repeatable associations across these two data sets, one of which was subsequently further validated in the MARC twinner population (Kirkpatrick, 2018: University of Wisconsin-Madison, Madison, WI, USA; unpublished). Two other regions with some degree of replication across studies are the region of bovine chromosome 23 containing the steroid 21-hydroxylase (CYP21) gene and chromosome 7 near the anti-Müllerian hormone (AMH) gene. In all three cases, contribution of these positional candidate genes to genetic variation is uncertain, and further work is needed to establish any causal relationship between these and variation in ovulation or twinning rate.

The search for major genes affecting ovulation in cattle led to the identification of the cow Treble with

an exceptional record of prolificacy (Morris *et al.*, 2010). This cow produced three sets of triplets in her lifetime among which was a son (Trio) who sired several daughters that had either twin or triplet births, suggesting transmission of a genetic factor across generations. Semen from Trio was used in artificial insemination at the University of Wisconsin-Madison and daughters born to these matings were evaluated for ovulation rate and genotypes for an initial within-family linkage analysis. The linkage analysis provided strong evidence ($P < 1 \times 10^{-28}$) of segregation of a single gene, located on chromosome 10, with a large effect on ovulation rate (1.02 ± 0.08 additional CL per cycle) (Kirkpatrick and Morris, 2015). Examination of this narrowed bovine genomic region suggested multiple candidate genes for the high ovulation rate phenotype. SMAD3 and SMAD6 reside in this region and are part of the TGF- β signaling system (ten Dijke and Hill, 2004). Considering the mediating roles of SMAD3 and SMAD6 in GDF9 and BMP15 signaling, respectively, and the high fecundity genotypes in sheep associated with GDF9 and BMP15, these two Smads were considered strong candidates for the causative mutation. However, screening for polymorphisms within the coding and flanking regions failed to identify the likely causative polymorphism (Kirkpatrick and Morris, 2015), suggesting that the functional polymorphism was either in a regulatory element located at a greater distance from the gene or is in a different gene in the region. Subsequent gene expression analyses in GC identified and confirmed significant overexpression of SMAD6 in Trio carrier cattle vs non-carrier, strongly implicating SMAD6 as the gene responsible for the high ovulation rate phenotype, though the actual mutation remains unknown (García-Guerra *et al.*, 2018a; Kamalludin *et al.*, 2018).

Table 2. Chromosomal locations of quantitative trait loci and single nucleotide polymorphisms associated with twinning rate and ovulation rate in cattle.

Trait	Chromosome and approximate location within chromosome (Mb)	Population	Positional candidate genes (chromosome)	Ref.
Ovulation rate	7 (40) and 23 (27)	MARC twinner	CYP21 (23)	Blattman <i>et al.</i> , 1996
Twinning rate	5 (64), 7 (108), 12 (10) and 23 (26)	Norwegian cattle	IGF1 (5), CYP21 (23)	Lien <i>et al.</i> , 2000; Meuwissen <i>et al.</i> , 2002
Ovulation rate	5 (46)	MARC twinner		Kappes <i>et al.</i> , 2000; Allan <i>et al.</i> , 2009
Twinning rate	5 (68)	US Holstein	IGF1 (5)	Cruickshank <i>et al.</i> , 2004; Kim <i>et al.</i> , 2009b
Twinning rate	8 (108), 10 (26) and 14 (51)	US Holstein		Cobanoglu <i>et al.</i> , 2005
Ovulation rate	14 (61)	MARC twinner		Gonda <i>et al.</i> , 2004
Ovulation rate	7 (22), 10 (75) and 19 (42)	MARC twinner	AMH (7), ESR2 (10), IGFBP4(19)	Arias and Kirkpatrick, 2004
Twinning rate	4 (44), 5 (67), 6 (8, 44), 7 (68, 76), 8 (58), 9 (34), 11 (47), 14 (21, 38), 15 (23), 23 (51), and 28 (9)	US Holstein	IGF1 (5)	Kim <i>et al.</i> , 2009a; Bierman <i>et al.</i> , 2010
Twinning rate	6 (51), 7 (19), 23 (27)		AMH (7), CYP21 (23)	Weller <i>et al.</i> , 2008
Twinning rate	24 (40)	Italian Maremmana		Moioli <i>et al.</i> , 2017



Physiology underlying multiple ovulation models

Ovine high fecundity

Mutations in high fecundity ovine genotypes, in general, result in reduced signaling of the GDF9 and/or BMP15 pathways leading to ovulation of multiple smaller follicles as compared to wild type ewes (McNatty *et al.*, 1986b). However, the overall ovarian phenotype depends on the precise mutation and the carrier status of the animal (heterozygous or homozygous). Table 1 shows a comprehensive list of the mutations identified in sheep and the associated ovarian phenotype (ovulation rate, percent increase, etc.) according to their carrier status. Homozygous carriers of high fecundity alleles show two distinct ovarian phenotypes. Approximately 60% of the mutations result in primary ovarian failure (POF) and an infertile phenotype in the homozygous state, due to block of follicle development at the primary or secondary stage. Similarly, immunization of ewes against GDF9 and BMP15 results, in most cases, in an anovulatory state and the presence of normal follicles up to the primary stage but only a few abnormal follicles developing past the primary stage (Juengel *et al.*, 2002; McNatty *et al.*, 2007). Conversely, immunization with some antigens, resulted initially in increased ovulation rate in some ewes. Taken together, these results indicate that a partial decrease in the availability of BMP15 and GDF9 underlies the occurrence of multiple ovulations.

On the other hand, some of the high fecundity ovine genotypes are fertile when present in the homozygous state. The Booroola/FecB results in a further increase in ovulation rate when present in the homozygous state, with ovulation rates that can reach up to 14, with homozygous ewes being fully fertile (Bindon and Piper, 1986; McNatty *et al.*, 2017). This mutation does not result in reduction in BMP1B mRNA or protein concentrations, but there is diminished receptor activity (Fabre *et al.*, 2006). More recently, mutations in BMP15 and GDF9 that do not result in POF and sterility in the homozygous state have been described (Table 1). These mutations have a further increase in ovulation rate when present in the homozygous state, although the increase is not as substantial as the one observed in Booroola (Silva *et al.*, 2011; Demars *et al.*, 2013). Furthermore, ewes with multiple fecundity mutations have substantial increases in ovulation rate indicating that they are independent and do not cancel each other out or lead to anovulatory phenotypes (Hanrahan *et al.*, 2004; Drouilhet *et al.*, 2009; McNatty *et al.*, 2017).

Among the models proposed to explain the occurrence of multiple ovulations in high fecundity ovine genotypes, one states that multiple ovulations could be the result of increasing the number of follicles available for selection (Baird and Campbell, 1998; Scaramuzzi *et al.*, 2011; Monniaux, 2016). Data on antral follicle counts (AFC) from ewes with or without high fecundity genes show considerable variation, with some studies indicating no differences in AFC while others indicate greater AFC in ewes carrying a high

fecundity allele. Booroola carrier ewes exhibit similar AFC (≥ 1 mm) as non-carriers as shown by multiple studies (McNatty *et al.*, 1985, 1986b; Henderson *et al.*, 1987; Gibbons *et al.*, 1999a). In agreement, no correlation between AFC and ovulation rate in the previous cycle was found in Booroola ewes (Driancourt *et al.*, 1985). However, a recent study showed lower circulating and intrafollicular AMH levels in Booroola/FecB homozygous ewes than wild type ewes but greater number of follicles ≥ 1 mm (Estienne *et al.*, 2015). In addition, the authors showed that BMP4-induced AMH production by GC was impaired in Booroola/FecB carriers. The authors postulated a model in which greater number of follicles coupled with reduced AMH production, could increase FSH and LH sensitivity and this would allow follicles to mature at a smaller size and in greater number (Estienne *et al.*, 2015).

Follicle populations in BMP15/FecX mutations have been the subject of fewer studies and there appears to be differences between specific lines. Heterozygous ewes for the Inverdale/FecX¹ mutations, were found to have greater AFC (≥ 1 and ≥ 2.5 mm) compared to wild type ewes (Shackell *et al.*, 1993; McNatty *et al.*, 2009). Conversely, in heterozygous Raza Aragonesa/FecX^R, another BMP15 mutation, no differences were found in AFC (≥ 3 mm) or circulating AMH when compared to controls (Lahoz *et al.*, 2013, 2014). Reports for other mutations are scarce in the literature, however, reports available on Ile de France/FecG^V and Lacaune/FecL^L ewes showed no differences in AFC between carrier and wild type ewes (Drouilhet *et al.*, 2010; Souza *et al.*, 2014). The role of antral follicle numbers on selection of multiple follicles has yet to be elucidated and further research is needed.

Several studies have conclusively demonstrated that ewes carrying high fecundity alleles such as Booroola/Fec^B (McNatty *et al.*, 1985, 1986b; Souza *et al.*, 1997), Inverdale/FecX¹ (Shackell *et al.*, 1993), Ile de France/FecG^V (Souza *et al.*, 2014), and Lacaune/FecL^L (Drouilhet *et al.*, 2010) have preovulatory follicles that are significantly smaller than those observed in non-carrier controls. Ewes carrying high fecundity alleles have fewer GC per follicle, compared to wild type control ewes even when comparing similar-sized follicles (McNatty *et al.*, 1985, 1986b, 2017). However, when number of GC in all presumptive preovulatory follicles were taken together, there were no differences in total GC number between wild types, Inverdale/FecX¹, homozygous or heterozygous Booroola/FecB, or even ewes carrying combinations of these mutations and Woodlands/FecW2^W (McNatty *et al.*, 1979, 1985, 1986b, 2017). Follicle dynamics have not been evaluated to a great extent between ewes carrying high fecundity alleles and wild type controls with the exception of two studies (Souza *et al.*, 1997; Gibbons *et al.*, 1999a). Both of these studies found smaller follicle sizes in high fecundity ewes starting 48 h after wave emergence.

In agreement with the smaller preovulatory follicle size, the resulting corpora lutea (CL) of ewes carrying high fecundity alleles (i.e. Booroola/FecB or



Inverdale FecX¹) are smaller on an individual basis and have fewer cells than those observed in non-carrier control ewes (McNatty *et al.*, 1985, 1986b, 2017; Niswender *et al.*, 1990; Shackell *et al.*, 1993). However, circulating P4, total luteal weight, luteal cell volume, number of cells/gr of tissue, and average cell dimensions were similar between genotypes (Niswender *et al.*, 1990; Souza *et al.*, 1997). Overall, high fecundity genotypes ovulate smaller follicles, leading to smaller individual CL, however due to the higher ovulation rate total luteal tissue and circulating P4 is similar.

Two independent studies showed that GC from Booroola/FecB carrier ewes have reduced proliferation compared to wild type ewes (Monniaux *et al.*, 2000; Mulsant *et al.*, 2001). Furthermore, treatment of GC with BMP4 increased thymidine uptake in wild-types while the same treatment applied to homozygous Booroola/FecB carriers was without effect (Mulsant *et al.*, 2001; Fabre *et al.*, 2003). Interestingly, the sera of ewes immunized against BMP15 or GDF9 reduced GC proliferation by 70% when sera originated from ewes that were anovular after immunization against BMP15 or GDF9. On the other hand, sera from animals with more than 3 CL inhibited thymidine incorporation by 24-27% (McNatty *et al.*, 2007). These results, taken together, provide some evidence that there is impaired signaling of the mutated BMPRII receptor in Booroola/FecB GC and that in order for multiple ovulations to occur GC proliferation needs to be only partially reduced.

Follicles of ewes carrying the Booroola mutation reached peak aromatase activity and follicular fluid E2 concentrations at progressively smaller diameters related to whether they were heterozygous or homozygous for the mutated allele (McNatty *et al.*, 1985, 1986b). Similar to E2, follicles from ewes carrying high fecundity alleles have GC that are responsive to LH stimulation at a smaller follicle size (McNatty *et al.*, 1986a, 2009, 2017), are capable of producing levels of cAMP, in response to LH, at smaller sizes (Henderson *et al.*, 1987), and have greater mRNA for LHCGR in GC of small (1-3 mm) and medium (3-4.5 mm) follicles than similar follicles from wild type controls (Juengel *et al.*, 2017). Thus, acquisition of LHCGR in GC, increased E2 production, and decreased IGFBP (Monniaux *et al.*, 2000) at smaller follicle sizes indicate that acquisition of dominance at a smaller follicle size is a hallmark of the physiology of ewes carrying high fecundity alleles.

It is well-known that exogenous FSH can override the selection mechanisms allowing for codominance and multiple ovulations in both sheep and cattle. Initial studies on circulating FSH in Booroola/FecB ewes found elevated mean concentrations in homozygous carriers, while heterozygous carriers were intermediate but not different from controls (McNatty *et al.*, 1987; Boulton *et al.*, 1995). Similarly, a recent study in ewes containing both the Booroola/FecB and Inverdale FecX¹, had greater circulating FSH on day 5-6 of the estrous cycle compared to controls (Juengel *et al.*, 2017). The differences found in circulating FSH

however do not appear to be associated with differences in circulating inhibin (McNatty *et al.*, 1992; Shackell *et al.*, 1993; Souza *et al.*, 1997). Conversely, other studies found no differences in circulating FSH between Booroola ewes (homozygous and heterozygous combined) and wild type controls (Souza *et al.*, 1997; Gibbons *et al.*, 1999a). The conflicting results in circulating FSH concentrations between carriers of the Booroola mutation and wild type ewes could potentially result from the use of different breed backgrounds, differences in assays, and finally different strategies to normalize FSH patterns between ewes. In this regard, only two studies evaluated circulating FSH normalized to emergence of the follicular wave and both failed to identify differences in FSH (Souza *et al.*, 1997; Gibbons *et al.*, 1999a). However, small follicle size and less pronounced diameter deviation in ewes make it reasonable to expect that subtle differences in FSH near follicle selection would go unnoticed.

One study that provided evidence against the role of changes in circulating FSH driving the occurrence of multiple ovulations in ovine, high-fecundity genotypes was done in carriers and non-carriers of Booroola FecB ewes using autotransplanted ovaries, a GnRH antagonist and controlled patterns of FSH and LH in both genotypes (Campbell *et al.*, 2003). Interestingly, the differences in ovulation rate between carriers and wild types were maintained despite similar FSH/LH patterns being delivered. The authors stated that these results did not support the idea that the FecB gene acts simply through increasing gonadotropin stimulation but that sensitivity of follicular cells to gonadotropins must also be critical for increased ovulation rate produced by the FecB allele. Thus, descriptive studies on FSH patterns support increases/alterations in circulating FSH in carriers of fecundity genotypes but definitive evidence has not yet been provided, whereas this manipulative study supports the idea that differences in circulating FSH may be secondary to ovarian gonadotropin sensitivity in determining the ovulation rate in carriers and non-carriers of fecundity genotypes.

USDA MARC twinner cattle

The twinner cattle population at USDA-MARC has been the result of selection over multiple generations for twinning and ovulation rate (Echternkamp *et al.*, 1990a; Gregory *et al.*, 1990). As a result, twinner cattle have on average 2.1 ovulations per cycle, with ~60% of the cycles having 2 ovulations and rarely exceeding 4 ovulations (Echternkamp *et al.*, 2009).

Follicle numbers have been investigated in MARC twinner cattle as a potential component of the mechanism underlying multiple ovulations, as previously described for high fecundity alleles in sheep. Total antral surface follicles are greater in twinner cows than in control cows from an unselected population (Echternkamp *et al.*, 1990b, 2004). Furthermore, differences have been found in the preantral follicle population, with twinner cows having significantly



more (~2-fold) secondary follicles than controls, while no differences were found in primordial, primary, or tertiary follicles (Cushman *et al.*, 2000). However, the control population used for comparison is of paramount importance as follicle numbers have been shown to be highly variable among individuals. The control population utilized in MARC twinner studies may have a different genetic makeup and this could potentially confound reported associations. Twinner cattle preovulatory follicle size (12 h after estrus) was also evaluated in relation to ovulation rate, indicating that individual follicle diameter decreased as ovulation rate increased from 1 to 3, but no further decrease was seen thereafter although sample size of cows with ovulation rates greater than 3 was low (Echternkamp *et al.*, 2009). Likewise, individual CL volume decreased with increased ovulation rate, however total CL volume, weight and circulating P4 increased with increasing ovulation rate (Echternkamp *et al.*, 2004, 2009). Circulating FSH in the USDA-MARC twinner population did not indicate differences when compared with unselected controls, although interpretation of the results is hampered by lack of normalization to wave emergence or deviation (Echternkamp, 2000; Echternkamp *et al.*, 2004). Thus, precise follicle dynamics and associated hormonal patterns of MARC twinner cattle, in relation to diameter deviation, have not yet been conclusively investigated.

Several aspects of the follicular microenvironment have been evaluated in relation to selection of multiple dominant follicles in the MARC twinner cattle population. The IGF1 system appears to be a key component of the altered selection mechanism in MARC twinner cows as evidenced by: 1) greater plasma and intrafollicular IGF1 (Echternkamp *et al.*, 1990b, 2004); 2) greater binding activity of IGFBP-3 and one form of IGFBP-5, but lower binding activity of one form of IGFBP-4 (Echternkamp *et al.*, 2004); and 3) decreased IGF2R mRNA in GC, an IGF2 receptor lacking kinase activity (Echternkamp *et al.*, 2012; Aad *et al.*, 2013). Thus, lower IGF2R and IGFBP-4 and greater IGF1 have been implicated in selection of additional follicles, although the precise sequence of events has not been resolved.

Trio, a novel high fecundity allele

The recent discovery of Trio provides an outstanding opportunity to investigate the physiologic mechanisms associated with selection of a single follicle (non-carriers) or multiple follicles (Trio carriers) during different experimental conditions (Kirkpatrick and Morris, 2015). The cow is a particularly useful research model for studies of follicular selection due to ease and accuracy of ultrasound evaluation of bovine follicular growth, ability to evaluate concurrent endocrine profiles, and methods to manipulate or evaluate follicular fluid (Ginther *et al.*, 1997b, 2004; Beg *et al.*, 2002; Beg and Ginther, 2006).

Evaluation of more than 243 estrous cycles in Trio carrier cattle (heterozygote) and half-sib non-carriers, revealed that Trio carriers had a consistent

increase in ovulation rate with a mean ovulation rate of 3.5 ± 0.2 while non-carriers had a mean ovulation rate of 1.1 ± 0.1 (García-Guerra *et al.*, 2017b). In Trio carriers, most (70.4%; 95/134) cycles had 3 or 4 ovulations, and few cycles had >5 ovulations (4.4%) or only a single ovulation (5.2%), while ~89% of the cycles of non-carrier controls had single ovulations, and no cycles were observed having 3 ovulations or more. Thus, heterozygous carriers of Trio have a dramatic and consistent increase in ovulation rate. More recently, ovulation rate was obtained from 3 homozygous Trio allele carriers during a minimum of 4 estrous cycles (García-Guerra *et al.*, 2018; University of Wisconsin-Madison, Madison, WI, USA; unpublished). Mean ovulation rate per cycle was 4.3 ± 0.5 for homozygous Trio allele carrier heifers, indicating that the Trio allele does not result in primary ovarian failure in the homozygous state, unlike many of the high fecundity genotypes found in sheep. However, due to low number of homozygote animals, it is currently unclear if homozygous Trio carriers have similar or increased ovulations compared to heterozygotes.

We recently investigated differences in antral follicle populations in carriers vs. non-carriers of Trio (García-Guerra *et al.*, 2017b). Trio carriers had AFC (≥ 2 mm) at the time of wave emergence that were similar to non-carriers. Furthermore, no association was observed between AFC and mean ovulation rate for the preceding four estrous cycles in Trio carriers or non-carriers despite large variation in AFC. Production of AMH in females is from GC of growing preantral and small antral follicles, thus providing a good representation of the dynamic follicle reserve (Rico *et al.*, 2011). In cattle, AMH and AFC have been found to have relatively high correlations ranging from 0.59 to 0.88 (Ireland *et al.*, 2008). As expected, our results indicate that circulating AMH concentrations were positively associated with AFC in both Trio carrier and non-carrier cattle (García-Guerra *et al.*, 2017b). However, there was no difference between genotypes in circulating AMH. Thus, differences in size of the antral follicle population is not causing selection of multiple follicles in cattle carrying the high fecundity allele, Trio.

Trio carriers exhibit predominantly (>70%) 3 follicular waves during the estrous cycle, similar to non-carriers (García-Guerra *et al.*, 2017a). In agreement with increased ovulation rate, each follicular wave contained greater number of dominant follicles in Trio carriers (3 to 4 per wave) compared to non-carriers (1 per wave). In addition, Trio carriers ovulate multiple smaller-sized follicles, however, the study of follicle dynamics in Trio carriers also indicates that follicles are smaller during the entire follicular wave (García-Guerra *et al.*, 2017a, 2018b). Evidence for smaller follicle size in Trio carriers arises from the following observations: 1) largest, smallest and mean preovulatory follicle diameter is greatly reduced; 2) future dominant follicles are smaller at wave emergence, at deviation, and at time of maximum diameter; 3) future first subordinate follicle is smaller at wave emergence and deviation; 4) reduced follicle growth rate; and 5) resulting CL are significantly smaller on an individual basis.

Nevertheless, there is clear evidence of subordinate follicles in Trio carriers, supporting the idea that selection still occurs but at a smaller size and with a greater number of selected follicles in Trio carriers. Thus, diameter deviation between future dominant follicles and subordinate follicles occurred at similar times (~3 days) in Trio carriers and non-carriers albeit at significantly smaller follicle sizes (6 mm vs. 8.6 mm, respectively).

Evaluation of CL volume and circulating P4 indicates that despite individual CL being smaller in Trio carriers, total luteal volume and resulting P4 concentrations are not different than those observed in non-carriers (García-Guerra *et al.*, 2017b, 2018b). As a result, it is reasonable to evaluate follicle size and growth patterns on a volume basis rather than diameter. As shown in Fig. 2, the diameter and volume of the largest or mean dominant follicle (average of all

dominant follicles) normalized to onset of diameter deviation is significantly smaller in Trio carriers compared to non-carrier controls (García-Guerra *et al.*, 2018b). However, total dominant follicle volume, calculated as the sum of each individual dominant follicle, is similar between genotypes. Interestingly, there is a close association between ovulation rate and individual follicle size such that Trio carrier cattle have ~4-fold greater number of ovulations than single-ovulating controls but their preovulatory follicles are individually ~4-fold smaller on a volume basis than in controls, thus the total preovulatory follicle volume is similar. In agreement, circulating E2, normalized to day of ovulation, is similar between genotypes (García-Guerra *et al.*, 2017a), indicating that combined hormonal output is similar regardless of whether it originates from one large follicle or four smaller follicles.

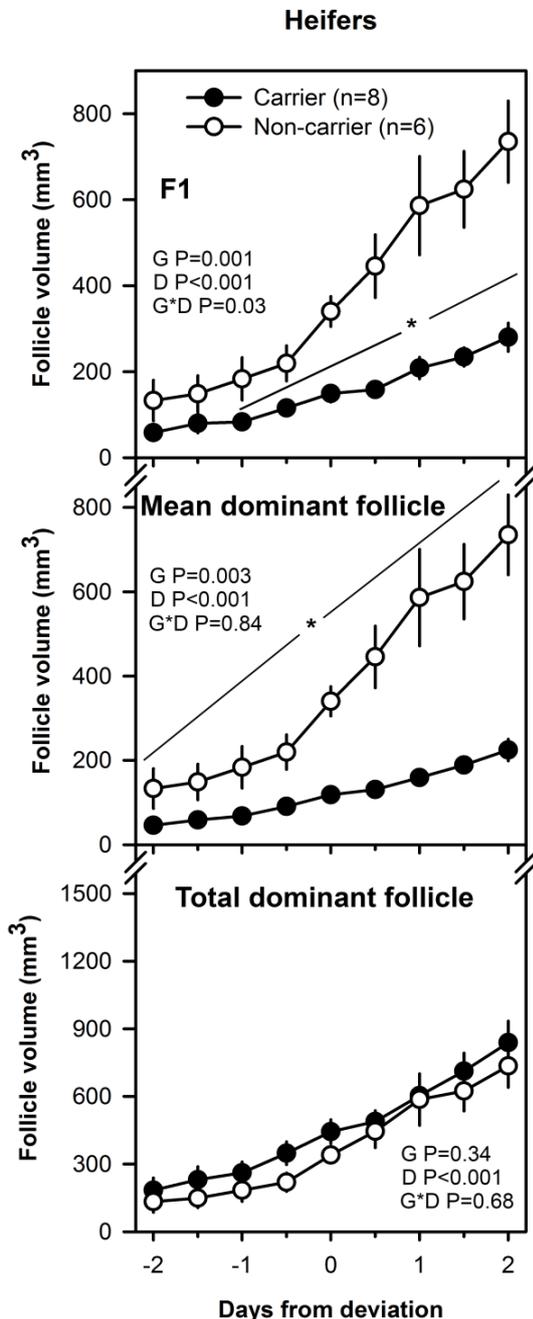


Figure 2. Growth profile of largest follicle volume (F1), mean dominant follicle volume, and total dominant follicle volume in Trio carrier and age-matched, half-sib non-carrier control heifers (adapted from García-Guerra *et al.*, 2017b). Data were normalized to deviation and each point represents mean (\pm SEM). *Indicates significant differences between genotype for a given time point ($P < 0.05$). G, genotype; D, day; G*D, genotype by day interaction.

The known role of FSH in stimulating follicle growth and the ability to induce multiple ovulations by administration of exogenous FSH (Adams *et al.*, 1993), leads naturally to the evaluation of FSH levels in selection of multiple dominant follicles in high fecundity genotypes. In this regard, FSH surges of similar magnitude were found to precede emergence of each follicular wave during the estrous cycle of Trio carriers and non-carriers; although circulating FSH was greater near follicular deviation (García-Guerra *et al.*, 2017a). In a second experiment, we synchronized the emergence of the follicular wave and evaluated circulating FSH at frequent intervals (García-Guerra *et al.*, 2018b). Overall, average circulating FSH was greater in Trio carriers than non-carriers. However, precise analysis focusing on the time encompassing observed deviation found that FSH was greater only in a narrow window encompassing deviation (Fig. 3). This elevation in circulating FSH concentrations near the time of follicular deviation is consistent with FSH being a key component of the process that allows selection of multiple dominant follicles in this high fecundity genotype. Nevertheless, provision of large doses of FSH produced a similar superovulatory response in either carriers or non-carriers of the Trio allele (García-Guerra *et al.*, 2018; University of Wisconsin-Madison, Madison, WI, USA; unpublished).

Based on the observation that diameter deviation occurs at a similar time but smaller follicle size in Trio carriers, we hypothesized that the dominant phenotype and ovulatory capacity also occurred at a smaller follicle size. We found that follicles of Trio carriers acquired a dominant phenotype, as determined by intrafollicular E2 concentrations, CYP19A1, LHCGR, and PAPP A mRNA abundance in GC, at a significantly smaller size than in non-carrier controls (García-Guerra *et al.*, 2018a). As expected, non-carrier single-ovulating cattle acquired a dominant phenotype when follicles were ~8.5 mm. Conversely, Trio carriers

acquired a similar phenotype when follicles were ~6 mm, thus supporting the idea that dominance is acquired at a smaller size and in agreement with observed diameter deviation. Moreover, in another experiment, challenge with an exogenous ovulatory stimulus revealed that ovulatory capacity (50% probability of ovulation) was achieved at a reduced follicle size in Trio carriers (5.5 mm) than non-carriers (8.3 mm). Moreover, when each factor pertaining to the dominant phenotype (i.e. E2, LHCGR) were analyzed individually, as a function of follicle size, the 50% probability of dominance was acquired at 5-5.5 mm and 8-8.5 mm for Trio carriers and non-carriers, respectively (García-Guerra *et al.*, 2018a). Consideration of follicle size on a volume basis further confirmed our previous findings indicating that acquisition of dominance occurred at a similar time but follicles of Trio carriers were approximately 1/3 to 1/4 the size, on a volume basis, compared to non-carriers.

Finally, based on timing of acquisition of the dominant phenotype, it appears follicles in Trio carriers acquire dominance in a hierarchal manner, as indicated by the increasing number of dominant follicles, based on intrafollicular E2, between days 2 and 4 after wave emergence (García-Guerra *et al.*, 2018a). Thus in Trio carriers, the first follicle to acquire dominance would be unable to provide sufficient FSH inhibitor (E2 and/or inhibin), due to its smaller size, to inhibit FSH thus the window for acquiring dominance remains open. As a result, the next follicle(s) in the hierarchy is (are) able to acquire a dominant phenotype and contribute to increasing circulating E2. The metaphorical gate of selection then, would remain open, until a sufficient number of dominant follicles (3 to 4) are present that equate to the same total follicle volume as that observed in a single dominant follicle in non-carrier cattle. This will produce sufficient FSH inhibitor (circulating E2) necessary to provide the final suppression of FSH and thus prevent selection of additional follicles.

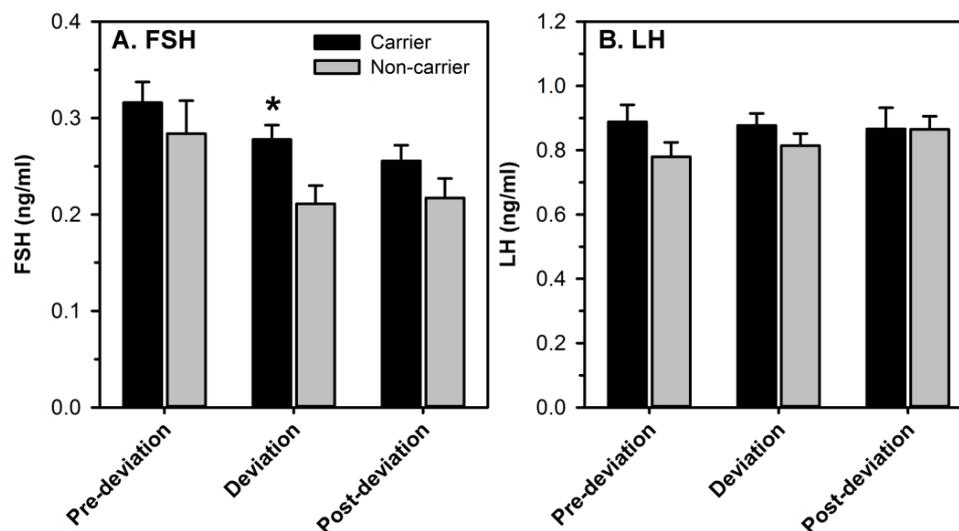


Figure 3. Mean FSH (A) and LH (B) concentrations at 3 distinct time points: pre-deviation (36 ± 12 h before the onset of deviation), at deviation (24 h encompassing the onset of deviation), and post-deviation (36 ± 12 h after the onset of deviation) in cattle with (Trio; n = 13) or without (Non-carriers; n = 9) the high fecundity allele (adapted from García-Guerra *et al.*, 2017b). Data represent mean (± SEM). * Indicates significant differences between genotypes (P < 0.05).

Conclusions - A model for selection of multiple follicles

Several models have been proposed to explain the occurrence of multiple ovulations in high fecundity genotypes in sheep, primarily based on studies of Booroola/FecB ewes (Baird 1987; Scaramuzzi *et al.*, 2011; Juengel *et al.*, 2013; Monniaux, 2016). Four potential, not mutually-exclusive, mechanisms have been proposed: 1) widening the selection gate by increasing FSH, which translates into a longer period of time during which FSH is above the threshold; 2) increasing the number of follicles, thus increasing the probability that more than one follicle is allowed through the gate; 3) decreasing GC proliferation and size at which follicles acquire LH receptors; and 4) increased sensitivity of follicles to FSH, thus allowing follicles to continue to grow in a lower FSH environment. Our recent data obtained from cattle carrying the high fecundity allele, Trio, are consistent with the first (slight FSH increase) and third (smaller follicles at dominance due to decreased GC proliferation) mechanisms but provide clear evidence against the second mechanism, increased number of follicles, as a mechanism to increase ovulation rate in Trio carriers. Although certain high fecundity ovine genotypes and the MARC-twiner bovine model have been reported to have greater numbers of small follicles, the mere increase in follicle numbers alone does not seem sufficient to explain the occurrence of increased multiple ovulations, since there is still no explanation for why only a portion of available follicles are selected. In addition, substantial variation in follicle numbers has been shown in single ovulating cattle, and no evidence has been provided so far to link this variation with the occurrence of multiple ovulations. Moreover, *Bos indicus* are known to have much greater numbers of antral follicles than *Bos taurus*, however the occurrence of multiple ovulations and births appears to be rare (Sartori and Barros, 2011). In a recent review (Monniaux, 2016), the increase in follicle numbers in regulation of multiple ovulations has been integrated with the fourth proposed mechanism (greater FSH sensitivity). The increase in FSH sensitivity has been hypothesized to arise from changes in number of FSH receptors, decreased AMH, increased response of the receptor upon stimulation, or increase in other factors that modulate or act in conjunction with FSH to stimulate GC (i.e. IGF1). Recent results from Trio carriers did not show an increased expression of FSH receptor or increased intrafollicular IGF1 but found decreased expression of both factors in dominant

follicles in Trio carriers as compared to controls (García-Guerra *et al.*, 2018a).

The most consistent finding for all high fecundity genotypes is smaller-sized dominant and ovulatory follicles in Trio carriers or ovine fecundity genotypes. In addition, our evidence in Trio, as well as evidence in high fecundity ovine genotypes support another aspect of the third mechanism, related to acquisition of LH receptors in smaller-sized follicles. Thus, development of smaller follicles that acquire dominance and ovulate at smaller sizes appears to be a key component for selection of multiple follicles. Thus, multiple smaller follicles function as a cohesive unit that has the same hormonal output (i.e. E2) and upon ovulation results in multiple CL of smaller individual size but the same total number of luteal cells with corresponding similar P4 output.

An important aspect that stems from the development of smaller follicles in Trio carriers is their reduced growth rate. The reduced follicle growth is likely mediated by a reduction in the rate at which GC proliferate, resulting from attenuation of BMP15 signaling pathways in response elevated SMAD6 (García-Guerra *et al.*, 2018a; Kamalludin *et al.*, 2018). Thus, overexpression of SMAD6 in GC of Trio carriers would be similar, functionally, to high-fecundity ovine mutations that affect BMP15 and GDF9. However, the precise effects of these factors in bovine GC along with the regulation of such pathways have not been investigated and should be the focus of future research.

The potential for FSH being the major driver for occurrence of multiple ovulations is extremely appealing. Our findings of elevated FSH concentrations, particularly around the time of expected deviation provides clear support for this mechanism, and this would provide a logical physiologic explanation for increased ovulation rate. Thus, our current model (Fig. 4) for Trio carriers is: 1) at wave emergence follicles are smaller (~50% of volume); 2) follicles develop at a reduced rate resulting in follicles ~25-30% of the volume of non-carriers at deviation; 3) upon reaching the decisive period (2.5-3 days) the largest follicle acquires dominance (i.e. LH receptors) with a corresponding increase in E2 production, however because of much smaller follicle size, the resulting E2 production is not sufficient to completely suppress FSH; 4) as a result the next follicle is allowed to acquire dominance (following a hierarchal order) until sufficient dominant follicle volume is reached, resulting in sufficient GC numbers to induce final suppression of FSH. Future studies are needed to definitely evaluate this model in other high fecundity genotypes.

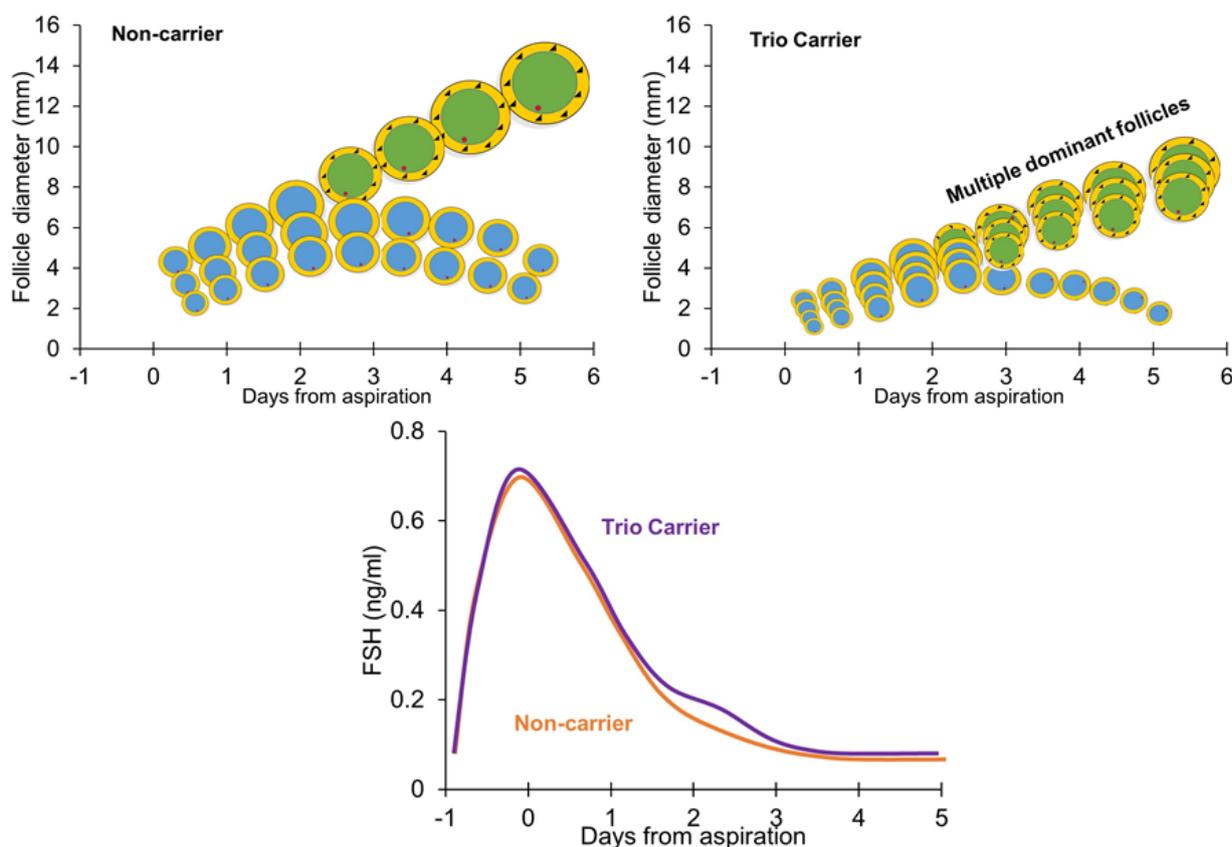


Figure 4. Physiological model for follicle selection in single ovulating cattle (left) and Trio carriers with multiple ovulations (right). The inset below shows the FSH concentrations during the period after follicle aspiration, emphasizing the comparison between carriers and non-carriers of the Trio allele.

References

- Aad PY, Echternkamp SE, Spicer LJ.** 2013. Possible role of IGF2 receptors in regulating selection of 2 dominant follicles in cattle selected for twin ovulations and births. *Domest Anim Endocrinol*, 45:187-195.
- Abdoli R, Zamani P, Deljou A, Rezvan H.** 2013. Association of BMPR-1B and GDF9 genes polymorphisms and secondary protein structure changes with reproduction traits in Mehraban ewes. *Gene*, 524:296-303.
- Adams GP, Kot K, Smith CA, Ginther OJ.** 1993. Effect of the dominant follicle on regression of its subordinates in heifers. *Can J Anim Sci*, 73:267-275.
- Adams GP.** 1994. Control of ovarian follicular wave dynamics in cattle: implications for synchronization & superstimulation. *Theriogenology*, 41:19-24.
- Allan MF, Kuehn LA, Cushman RA, Snelling WM, Echternkamp SE, Thallman RM.** 2009. Confirmation of quantitative trait loci using a low-density single nucleotide polymorphism map for twinning and ovulation rate on bovine chromosome 5. *J Anim Sci*, 87:46-56.
- Arias J, Kirkpatrick B.** 2004. Mapping of bovine ovulation rate QTL; an analytical approach for three generation pedigrees. *Anim Genet*, 35:7-13.
- Baird DT.** 1987. A model for follicular selection and ovulation: Lessons from superovulation. *J Steroid Biochem*, 27:15-23.
- Baird DT, Campbell BK.** 1998. Follicle selection in sheep with breed differences in ovulation rate. *Mol Cell Endocrinol*, 145:89-95.
- Beg MA, Bergfelt DR, Kot K, Wiltbank MC, Ginther OJ.** 2000. Follicle selection in cattle: differential changes in intrafollicular concentrations of estradiol and insulin-like growth factor binding protein-2. *Biol Reprod*, 62:218-219.
- Beg MA, Bergfelt DR, Kot K, Wiltbank MC, Ginther OJ.** 2001. Follicular-fluid factors and granulosa-cell gene expression associated with follicle deviation in cattle. *Biol Reprod*, 64:432-441.
- Beg MA, Bergfelt DR, Kot K, Ginther OJ.** 2002. Follicle selection in cattle: dynamics of follicular fluid factors during development of follicle dominance. *Biol Reprod*, 66:120-126.
- Beg MA, Meira C, Bergfelt DR, Ginther OJ.** 2003. Role of oestradiol in growth of follicles and follicle deviation in heifers. *Reproduction*, 125:847-854.
- Beg MA, Ginther OJ.** 2006. Follicle selection in cattle and horses: role of intrafollicular factors. *Reproduction*, 132:365-377.
- Bergfelt DR, Kulick LJ, Kot K, Ginther OJ.** 2000. Follicular and hormonal response to experimental suppression of FSH during follicle deviation in cattle. *Theriogenology*, 54:1191-1206.
- Bertoldo MJ, Walters KA, Ledger WL, Gilchrist RB, Mermillod P, Locatelli Y.** 2018. In-vitro regulation of primordial follicle activation: challenges



- for fertility preservation strategies. *Reprod Biomed Online*, 10.1016/j.rbmo.2018.01.014.
- Bierman CD, Kim E, Shi XW, Weigel K, Jeffrey Berger P, Kirkpatrick BW.** 2010. Validation of whole genome linkage-linkage disequilibrium and association results, and identification of markers to predict genetic merit for twinning. *Anim Genet*, 41:406-416.
- Bindon BM.** 1984. Reproductive biology of the Booroola Merino sheep. *Aust J Biol Sci*, 37:163-189.
- Bindon BM, Piper LR.** 1986. Booroola (F) gene: major gene affecting ovine ovarian function. *Basic Life Sci*, 37:67-93.
- Blattman AN, Kirkpatrick BW, Gregory KE.** 1996. A search for quantitative trait loci for ovulation rate in cattle. *Anim Genet*, 27:157-162.
- Bodin L, Pasquale ED, Fabre S, Bontoux M, Monget P, Persani L, Mulsant P.** 2007. A novel mutation in the bone morphogenetic protein 15 gene causing defective protein secretion is associated with both increased ovulation rate and sterility in Lacaune sheep. *Endocrinology*, 148:393-400.
- Boulton MI, Haley CS, Springbett AJ, Webb R.** 1995. The effect of the Booroola (FecB) gene on peripheral FSH concentrations and ovulation rates during estrus, seasonal anestrus and on FSH concentrations following ovariectomy in Scottish-Blackface ewes. *J Reprod Fertil*, 103:199-207.
- Braw-Tal R, McNatty KP, Smith P, Heath DA, Hudson NL, Phillips DJ, McLeod BJ, Davis GH.** 1993. Ovaries of ewes homozygous for the X-linked Inverdale gene (FecX^L) are devoid of secondary and tertiary follicles but contain many abnormal structures. *Biol Reprod*, 49:895-907.
- Campbell BK, Baird DT, Souza CJ, Webb R.** 2003. The FecB (Booroola) gene acts at the ovary: in vivo evidence. *Reproduction*, 126:101-111.
- Cobanoglu O, Berger PJ, Kirkpatrick BW.** 2005. Genome screen for twinning rate QTL in four North American Holstein families. *Anim Genet*, 36:303-308.
- Cruickshank J, Dentine MR, Berger PJ, Kirkpatrick BW.** 2004. Evidence for quantitative trait loci affecting twinning rate in North American Holstein cattle. *Anim Genet*, 35:206-212.
- Cushman RA, Hedgpeth VS, Echterkamp SE, Britt JH.** 2000. Evaluation of numbers of microscopic and macroscopic follicles in cattle selected for twinning. *J Anim Sci*, 78:1564-1567.
- Davis GH, Montgomery GW, Allison AJ, Kelly RW, Bray AR.** 1982. Segregation of a major gene influencing fecundity in progeny of Booroola sheep. *New Zeal J Agr Res*, 25:525-529.
- Davis GH, Bruce GD, Dodds KG.** 2001a. Ovulation rate and litter size of prolific Inverdale (FecX^L) and Hanna (FecX^H) sheep. *Proc Assoc Advmt Anim Breed Genet*, 14:175-178.
- Davis GH, Dodds KG, Wheeler R, Jay NP.** 2001b. Evidence that an imprinted gene on the X chromosome increases ovulation rate in sheep. *Biol Reprod*, 64:216-221.
- Davis GH, Farquhar PA, O'Connell AR, Everett-Hincks JM, Wishart PJ, Galloway SM, Dodds KG.** 2006. A putative autosomal gene increasing ovulation rate in Romney sheep. *Anim Reprod Sci*, 92:65-73.
- Demars J, Fabre S, Sarry J, Rossetti R, Gilbert H, Persani L, Tosser-Klopp G, Mulsant P, Nowak Z, Drobik W, Martyniuk E, Bodin L.** 2013. Genome-Wide association studies identify two novel BMP15 mutations responsible for an atypical hyperprolificacy phenotype in sheep. *PLoS Genet*, 9:e1003482. doi: 10.1371/journal.pgen.1003482.
- Driancourt MA, Cahill LP, Bindon BM.** 1985. Ovarian follicular populations and preovulatory enlargement in Booroola and control Merino ewes. *J Reprod Fertil*, 73:93-107.
- Drouilhet L, Lecerf F, Bodin L, Fabre S, Mulsant P.** 2009. Fine mapping of the FecL locus influencing prolificacy in Lacaune sheep. *Anim Genet*, 40:804-812.
- Drouilhet L, Taragnat C, Fontaine J, Duittoz A, Mulsant P, Bodin L, Fabre S.** 2010. Endocrine characterization of the reproductive axis in highly prolific lacaune sheep homozygous for the FecLL mutation. *Biol Reprod*, 82:815-824.
- Echterkamp SE, Gregory KE, Dickerson GE, Cundiff LV, Koch RM, Van Vleck LD.** 1990a. Twinning in cattle: II. Genetic and environmental effects on ovulation rate in puberal heifers and postpartum cows and the effects of ovulation rate on embryonic survival. *J Anim Sci*, 68:1877-1888.
- Echterkamp SE, Spicer LJ, Gregory KE, Canning SF, Hammond JM.** 1990b. Concentrations of insulin-like growth factor-I in blood and ovarian follicular fluid of cattle selected for twins. *Biol Reprod*, 43:8-14.
- Echterkamp SE.** 2000. Endocrinology of increased ovarian folliculogenesis in cattle selected for twin births. *J Anim Sci*, 77:1-20.
- Echterkamp SE, Roberts AJ, Lunstra DD, Wise T, Spicer LJ.** 2004. Ovarian follicular development in cattle selected for twin ovulations and births. *J Anim Sci*, 82:459-471.
- Echterkamp SE, Cushman RA, Allan MF.** 2009. Size of ovulatory follicles in cattle expressing multiple ovulations naturally and its influence on corpus luteum development and fertility. *J Anim Sci*, 87:3556-3568.
- Echterkamp SE, Aad PY, Eborn DR, Spicer LJ.** 2012. Increased abundance of aromatase and follicle stimulating hormone receptor mRNA and decreased insulin-like growth factor-2 receptor mRNA in small ovarian follicles of cattle selected for twin births. *J Anim Sci*, 90:2193-2200.
- Estienne A, Pierre A, di Clemente N, Picard JY, Jarrier P, Mansanet C, Monniaux D, Fabre S.** 2015. Anti-Mullerian hormone regulation by the bone morphogenetic proteins in the sheep ovary: deciphering a direct regulatory pathway. *Endocrinology*, 156:301-313.
- Fabre S, Pierre A, Pisselet C, Mulsant P, Lecerf F, Pohl J, Monget P, Monniaux D.** 2003. The Booroola mutation in sheep is associated with an alteration of the bone morphogenetic protein receptor-IB functionality. *J Endocrinol*, 177:435-444.
- Fabre S, Pierre A, Mulsant P, Bodin L, Di Pasquale E, Persani L, Monget P, Monniaux D.** 2006. Regulation of ovulation rate in mammals: contribution of sheep genetic models. *Reprod Biol Endocrinol*, 4:20. doi: 10.1186/1477-7827-4-20.



- Fike KE, Bergfeld EG, Cupp AS, Kojima FN, Mariscal V, Sanchez T, Wehrman ME, Grotjan HE, Hamernik DL, Kittok RJ, Kinder JE. 1997. Gonadotropin secretion and development of ovarian follicles during oestrous cycles in heifers treated with luteinizing hormone releasing hormone antagonist. *Anim Reprod Sci*, 49:83-100.
- Galloway SM, McNatty KP, Cambridge LM, Laitinen MP, Juengel JL, Jokiranta TS, McLaren RJ, Luoro K, Dodds KG, Montgomery GW, Beattie AE, Davis GH, Ritvos O. 2000. Mutations in an oocyte-derived growth factor gene (BMP15) cause increased ovulation rate and infertility in a dosage-sensitive manner. *Nat Genet*, 25:279-283.
- García-Guerra A, Kirkpatrick BW, Wiltbank MC. 2017a. Follicular waves and hormonal profiles during the estrous cycle of carriers and non-carriers of the Trio allele, a major bovine gene for high ovulation and fecundity. *Theriogenology*, 100:100-113.
- García-Guerra A, Motta JCL, Melo LF, Kirkpatrick BW, Wiltbank MC. 2017b. Ovulation rate, antral follicle count, and circulating anti-Müllerian hormone in Trio allele carriers, a novel high fecundity bovine genotype. *Theriogenology*, 101:81-90.
- García-Guerra A, Canavessi AMO, Monteiro PLJ, Mezera MA, Sartori R, Kirkpatrick BW, Wiltbank MC. 2018a. Trio, a novel high fecundity allele: III. Acquisition of dominance and ovulatory capacity at a smaller follicle size. *Biol Reprod*, 98:350-365.
- García-Guerra A, Kamalludin MH, Kirkpatrick BW, Wiltbank MC. 2018b. Trio, a bovine high fecundity allele: II. Hormonal profile and follicular dynamics underlying the high ovulation rate. *Biol Reprod*, 98:335-349.
- Gasperin BG, Ferreira R, Rovani MT, Santos JT, Buratini J, Price CA, Gonçalves PBD. 2012. FGF10 inhibits dominant follicle growth and estradiol secretion in vivo in cattle. *Reproduction*, 143:815-823.
- Gibbons JR, Wiltbank MC, Ginther OJ. 1997. Functional interrelationships between follicles greater than 4 mm and the follicle-stimulating hormone surge in heifers. *Biol Reprod*, 57:1066-1073.
- Gibbons JR, Kot K, Thomas DL, Wiltbank MC, Gilther OJ. 1999a. Follicular and FSH dynamics in ewes with a history of high and low ovulation rates. *Theriogenology*, 52:1005-1020.
- Gibbons JR, Wiltbank MC, Ginther OJ. 1999b. Relationship between follicular development and the decline in the follicle-stimulating hormone surge in heifers. *Biol Reprod*, 60:72-77.
- Gilchrist RB, Ritter LJ, Myllymaa S, Kaivo-Oja N, Dragovic RA, Hickey TE, Ritvos O, Mottershead DG. 2006. Molecular basis of oocyte-paracrine signalling that promotes granulosa cell proliferation. *J Cell Sci*, 119:3811-3821.
- Gimenes LU, Sa MF, Carvalho NAT, Torres JRS, Souza AH, Madureira EH, Trinca LA, Sartorelli ES, Barros CM, Carvalho JBP, Mapletoft RJ, Baruselli PS. 2008. Follicle deviation and ovulatory capacity in *Bos indicus* heifers. *Theriogenology*, 69:852-858.
- Ginther OJ, Kastelic JP, Knopf L. 1989. Composition and characteristics of follicular waves during the bovine estrous cycle. *Anim Reprod Sci*, 20:187-200.
- Ginther OJ, Kot K, Wiltbank MC. 1995. Associations between emergence of follicular waves and fluctuations in FSH concentrations during the estrous-cycle in ewes. *Theriogenology*, 43:689-703.
- Ginther OJ, Wiltbank MC, Fricke PM, Gibbons JR, Kot K. 1996. Selection of the dominant follicle in cattle. *Biol Reprod*, 55:1187-1194.
- Ginther OJ, Kot K, Kulick LJ, Wiltbank MC. 1997a. Emergence and deviation of follicles during the development of follicular waves in cattle. *Theriogenology*, 48:75-87.
- Ginther OJ, Kot K, Kulick LJ, Wiltbank MC. 1997b. Sampling follicular fluid without altering follicular status in cattle: Oestradiol concentrations early in a follicular wave. *J Reprod Fertil*, 109:181-186.
- Ginther OJ, Bergfelt DR, Kulick LJ, Kot K. 1999. Selection of the dominant follicle in cattle: establishment of follicle deviation in less than 8 hours through depression of FSH concentrations. *Theriogenology*, 52:1079-1093.
- Ginther OJ. 2000. The FSH-follicle coupling hypothesis for follicle selection. *Biol Reprod*, 62:92-92.
- Ginther OJ, Bergfelt DR, Kulick LJ, Kot K. 2000a. Selection of the dominant follicle in cattle: role of estradiol. *Biol Reprod*, 63:383-389.
- Ginther OJ, Bergfelt DR, Kulick LJ, Kot K. 2000b. Selection of the dominant follicle in cattle: role of two-way functional coupling between follicle-stimulating hormone and the follicles. *Biol Reprod*, 62:920-927.
- Ginther OJ, Beg MA, Bergfelt DR, Donadeu FX, Kot K. 2001. Follicle selection in monovular species. *Biol Reprod*, 65:638-647.
- Ginther OJ, Bergfelt DR, Beg MA, Meira C, Kot K. 2004. In vivo effects of an intrafollicular injection of insulin-like growth factor I on the mechanism of follicle deviation in heifers and mares. *Biol Reprod*, 70:99-105.
- Ginther OJ, Siddiqui MAR, Baldrighi JM, Wolf CA, Castro T. 2016. Temporality of two-way functional coupling between FSH and follicles in heifers. *Theriogenology*, 86:1645-1653.
- Gonda MG, Arias JA, Shook GE, Kirkpatrick BW. 2004. Identification of an ovulation rate QTL in cattle on BTA14 using selective DNA pooling and interval mapping. *Anim Genet*, 35:298-304.
- Goto K, Kamiya Y, Imamura T, Miyazono K, Miyazawa K. 2007. Selective inhibitory effects of Smad6 on bone morphogenetic protein type I receptors. *J Biol Chem*, 282:20603-20611.
- Gregory KE, Echterkamp SE, Dickerson GE, Cundiff LV, Koch RM, Van Vleck LD. 1990. Twinning in cattle: I. Foundation animals and genetic and environmental effects on twinning rate. *J Anim Sci*, 68:1867-1876.
- Hanrahan JP, Gregan SM, Mulsant P, Mullen M, Davis GH, Powell R, Galloway SM. 2004. Mutations in the genes for oocyte-derived growth factors GDF9 and BMP15 are associated with both increased ovulation rate and sterility in Cambridge and Belclare sheep (*Ovis aries*). *Biol Reprod*, 70:900-909.
- Hata A, Lagna G, Massague J, Hemmati-Brivanlou A. 1998. Smad6 inhibits BMP/Smad1 signaling by



- specifically competing with the Smad4 tumor suppressor. *Genes Dev*, 12:186-197.
- Haughian JM, Ginther OJ, Diaz FJ, Wiltbank MC.** 2013. Gonadotropin-releasing hormone, estradiol, and inhibin regulation of follicle-stimulating hormone and luteinizing hormone surges: implications for follicle emergence and selection in heifers. *Biol Reprod*, 88:165. doi: 10.1095/biolreprod.112.107342.
- Heath DA, Pitman JL, McNatty KP.** 2017. Molecular forms of ruminant BMP15 and GDF9 and putative interactions with receptors. *Reproduction*, 154:521-534.
- Henderson KM, McNatty KP, Okeeffe LE, Lun S, Heath DA, Prisk MD.** 1987. Differences in gonadotropin-stimulated cyclic-AMP production by granulosa-cells from Booroola x Merino ewes which were homozygous, heterozygous or noncarriers of a fecundity gene influencing their ovulation rate. *J Reprod Fertil*, 81:395-402.
- Hsueh AJ, Kawamura K, Cheng Y, Fauser BC.** 2015. Intraovarian control of early folliculogenesis. *Endocr Rev*, 36:1-24. doi: 10.1210/er.2014-1020.
- Imamura T, Takase M, Nishihara A, Oeda E, Hanai J-i, Kawabata M, Miyazono K.** 1997. Smad6 inhibits signalling by the TGF- β superfamily. *Nature*, 389:622-626.
- Ireland JLH, Scheetz D, Jimenez-Krassel F, Themmen APN, Ward F, Lonergan P, Smith GW, Perez GI, Evans ACO, Ireland JJ.** 2008. Antral follicle count reliably predicts number of morphologically healthy oocytes and follicles in ovaries of young adult cattle. *Biol Reprod*, 79:1219-1225.
- Juengel JL, Hudson NL, Heath DA, Smith P, Reader KL, Lawrence SB, O'Connell AR, Laitinen MPE, Cranfield M, Groome NP, Ritvos O, McNatty KP.** 2002. Growth differentiation factor 9 and bone morphogenetic protein 15 are essential for ovarian follicular development in sheep. *Biol Reprod*, 67:1777-1789.
- Juengel JL, O'Connell AR, French MC, Proctor LE, Wheeler R, Farquhar PA, Dodds KG, Galloway SM, Johnstone PD, Davis GH.** 2011. Identification of a line of sheep carrying a putative autosomal gene increasing ovulation rate in sheep that does not appear to interact with mutations in the transforming growth factor beta superfamily. *Biol Reprod*, 85:113-120.
- Juengel JL, Davis GH, McNatty KP.** 2013. Using sheep lines with mutations in single genes to better understand ovarian function. *Reproduction*, 146:R111-R123.
- Juengel JL, French MC, Quirke LD, Kauff A, Smith GW, Johnstone PD.** 2017. Differential expression of CART in ewes with differing ovulation rates. *Reproduction*, 153:471-479.
- Kallen A, Polotsky AJ, Johnson J.** 2018. Untapped reserves: controlling primordial follicle growth activation. *Trends Mol Med*, 24:319-331.
- Kamalludin MH, Garcia-Guerra A, Wiltbank M, Kirkpatrick BW.** 2018. Trio, a novel high fecundity allele: I. Transcriptome analysis of granulosa cells from carriers and non-carriers of a major gene for bovine ovulation rate. *Biol Reprod*, 98:323-334.
- Kaneko H, Nakanishi Y, Taya K, Kishi H, Watanabe G, Sasamoto S, Hasegawa Y.** 1993. Evidence that inhibin is an important factor in the regulation of FSH-secretion during the mid-luteal phase in cows. *J Endocrinol*, 136:35-41.
- Kappes SM, Bennett GL, Keele JW, Echterkamp SE, Gregory KE, Thallman RM.** 2000. Initial results of genomic scans for ovulation rate in a cattle population selected for increased twinning rate. *J Anim Sci*, 78:3053-3059.
- Kim ES, Berger PJ, Kirkpatrick BW.** 2009a. Genome-wide scan for bovine twinning rate QTL using linkage disequilibrium. *Anim Genet*, 40:300-307.
- Kim ES, Shi X, Cobanoglu O, Weigel K, Berger PJ, Kirkpatrick BW.** 2009b. Refined mapping of twinning-rate quantitative trait loci on bovine chromosome 5 and analysis of insulin-like growth factor-1 as a positional candidate gene. *J Anim Sci*, 87:835-843.
- Kirkpatrick BW, Morris CA.** 2015. A major gene for bovine ovulation rate. *PLoS ONE*, 10:e0129025.
- Knopf L, Kastelic JP, Schallenberger E, Ginther OJ.** 1989. Ovarian follicular dynamics in heifers - test of 2-wave hypothesis by ultrasonically monitoring individual follicles. *Domest Anim Endocrinol*, 6:111-119.
- Kulick LJ, Kot K, Wiltbank MC, Ginther OJ.** 1999. Follicular and hormonal dynamics during the first follicular wave in heifers. *Theriogenology*, 52:913-921.
- Lahoz B, Alabart JL, Jurado JJ, Calvo JH, Martinez-Royo A, Fantova E, Folch J.** 2011. Effect of the FecX(R) polymorphism in the bone morphogenetic protein 15 gene on natural or equine chorionic gonadotropin-induced ovulation rate and litter size in Rasa Aragonesa ewes and implications for on-farm application. *J Anim Sci*, 89:3522-3530.
- Lahoz B, Alabart JL, Folch J, Sánchez P, Echegoyen E, Cocero MJ.** 2013. Influence of the FecX^R allele in heterozygous ewes on follicular population and outcomes of IVP and ET using LOPU-derived oocytes. *Reprod Domest Anim*, 48:717-723.
- Lahoz B, Alabart JL, Cocero MJ, Monniaux D, Echegoyen E, Sanchez P, Folch J.** 2014. Anti-Mullerian hormone concentration in sheep and its dependence of age and independence of BMP15 genotype: an endocrine predictor to select the best donors for embryo biotechnologies. *Theriogenology*, 81:347-357.
- Lassoued N, Benkhilil Z, Woloszyn F, Rejeb A, Aouina M, Rezik M, Fabre S, Bedhiaf-Romdhani S.** 2017. FecX (Bar) a Novel BMP15 mutation responsible for prolificacy and female sterility in Tunisian Barbarine sheep. *BMC Genet*, 18:43.
- Li Q, Agno JE, Edson MA, Nagaraja AK, Nagashima T, Matzuk MM.** 2011. Transforming growth factor β receptor type 1 is essential for female reproductive tract integrity and function. *PLoS Genet*, 7:e1002320. doi: 10.1371/journal.pgen.1002320.
- Li Q.** 2015. Inhibitory SMADs: potential regulators of ovarian function. *Biol Reprod*, 92:50. doi: 10.1095/biolreprod.114.125203.
- Liao WX, Moore RK, Otsuka F, Shimasaki S.** 2003. Effect of intracellular interactions on the processing and secretion of bone morphogenetic protein-15 (BMP-15)



- and growth and differentiation factor-9: Implication of the aberrant ovarian phenotype of BMP15 mutant sheep. *J Biol Chem*, 278:3713-3719.
- Lien S, Karlsten A, Klemetsdal G, Vage DI, Olsaker I, Klungland H, Aasland M, Heringstad B, Ruane J, Gomez-Raya L.** 2000. A primary screen of the bovine genome for quantitative trait loci affecting twinning rate. *Mamm Genome*, 11:877-882.
- Lopez H, Sartori R, Wiltbank MC.** 2005. Reproductive hormones and follicular growth during development of one or multiple dominant follicles in cattle. *Biol Reprod*, 72:788-795.
- Luo W, Piccinato CA, Wiltbank MC.** 2003. Differential timing in cAMP regulation of cyclins, aromatase, and LH receptor mRNA in cultured bovine granulosa cells from 5 mm follicles. *Biol Reprod*, (suppl):114. (abstract).
- Luo W, Gumen A, Haughian JM, Wiltbank MC.** 2011. The role of luteinizing hormone in regulating gene expression during selection of a dominant follicle in cattle. *Biol Reprod*, 84:369-378.
- Martinez-Royo A, Jurado JJ, Smulders JP, Marti JJ, Alabart JL, Roche A, Fantova E, Bodin L, Mulsant P, Serrano M, Folch J, Calvo JH.** 2008. A deletion in the bone morphogenetic protein 15 gene causes sterility and increased prolificacy in Rasa Aragonesa sheep. *Anim Genet*, 39:294-297.
- McNatty KP, Smith DM, Makris A, Osathanondh R, Ryan KJ.** 1979. The microenvironment of the human antral follicle: Interrelationships among the steroid levels in antral fluid, the population of granulosa cells, and the status of the oocyte in vivo and in vitro. *J Clin Endocrinol Metab*, 49:851-860.
- McNatty KP, Henderson KM, Lun S, Heath DA, Ball K, Hudson NL, Fannin J, Gibb M, Kieboom LE, Smith P.** 1985. Ovarian activity in Booroola × Romney ewes which have a major gene influencing their ovulation rate. *J Reprod Fertil*, 73:109-120.
- McNatty KP, Kieboom LE, McDiarmid J, Heath DA, Lun S.** 1986a. Adenosine cyclic 3'-monophosphate and steroid production by small ovarian follicles from Booroola ewes with and without a fecundity gene. *J Reprod Fertil*, 76:471-480.
- McNatty KP, Lun S, Heath DA, Ball K, Smith P, Hudson NL, McDiarmid J, Gibb M and Henderson KM.** 1986b. Differences in ovarian activity between Booroola × Merino ewes which were homozygous, heterozygous and non-carriers of a major gene influencing their ovulation rate. *J Reprod Fertil*, 77:193-205.
- McNatty KP, Hudson N, Henderson KM, Gibb M, Morrison L, Ball K, Smith P.** 1987. Differences in gonadotropin concentrations and pituitary-responsiveness to GnRH between Booroola ewes which were homozygous (FF), heterozygous (F+), and noncarriers (++) of a major gene influencing their ovulation rate. *J Reprod Fertil*, 80:577-588.
- McNatty KP, Heath DA, Hudson NL, Ball K, Condell L.** 1992. Concentrations of immunoreactive inhibin in ovarian and peripheral venous plasma and follicular fluid of Booroola ewes that are homozygous carriers or non-carriers of the FecB gene. *J Reprod Fertil*, 95:489-502.
- McNatty KP, Moore LG, Hudson NL, Quirke LD, Lawrence SB, Reader K, Hanrahan JP, Smith P, Groome NP, Laitinen M, Ritvos O, Juengel JL.** 2004. The oocyte and its role in regulating ovulation rate: a new paradigm in reproductive biology. *Reproduction*, 128:379-386.
- McNatty KP, Juengel JL, Reader KL, Lun S, Myllymaa S, Lawrence SB, Western A, Meerasahib MF, Mottershead DG, Groome NP, Ritvos O, Laitinen MPE.** 2005a. Bone morphogenetic protein 15 and growth differentiation factor 9 co-operate to regulate granulosa cell function. *Reproduction*, 129:473-480.
- McNatty KP, Juengel JL, Reader KL, Lun S, Myllymaa S, Lawrence SB, Western A, Meerasahib MF, Mottershead DG, Groome NP, Ritvos O, Laitinen MPE.** 2005b. Bone morphogenetic protein 15 and growth differentiation factor 9 co-operate to regulate granulosa cell function in ruminants. *Reproduction*, 129:481-487.
- McNatty KP, Hudson NL, Whiting L, Reader KL, Lun S, Western A, Heath DA, Smith P, Moore LG, Juengel JL.** 2007. The effects of immunizing sheep with different BMP15 or GDF9 peptide sequences on ovarian follicular activity and ovulation rate. *Biol Reprod*, 76:552-560.
- McNatty KP, Heath DA, Hudson NL, Lun S, Juengel JL, Moore LG.** 2009. Gonadotrophin-responsiveness of granulosa cells from bone morphogenetic protein 15 heterozygous mutant sheep. *Reproduction*, 138:545-551.
- McNatty KP, Heath DA, Clark Z, Reader K, Juengel JL, Pitman JL.** 2017. Ovarian characteristics in sheep with multiple fecundity genes. *Reproduction*, 153:233-240.
- Meuwissen TH, Karlsten A, Lien S, Olsaker I, Goddard ME.** 2002. Fine mapping of a quantitative trait locus for twinning rate using combined linkage and linkage disequilibrium mapping. *Genetics*, 161:373-379.
- Moioli B, Steri R, Marchitelli C, Catillo G, Buttazzoni L.** 2017. Genetic parameters and genome-wide associations of twinning rate in a local breed, the Maremmana cattle. *Animal*, 11:1660-1666.
- Monniaux D, Monget P, Pisselet C, Fontaine J, Elsen JM.** 2000. Consequences of the presence of the Booroola F gene on the intraovarian insulin-like growth factor system and terminal follicular maturation in Mérinos d'Arles ewes. *Biol Reprod*, 63:1205-1213.
- Monniaux D.** 2016. Driving folliculogenesis by the oocyte-somatic cell dialog: Lessons from genetic models. *Theriogenology*, 86:41-53.
- Moore RK, Otsuka F, Shimasaki S.** 2003. Molecular basis of bone morphogenetic protein-15 signaling in granulosa cells. *J Biol Chem*, 278:304-310.
- Morris CA, Wheeler M, Levet GL, Kirkpatrick BW.** 2010. A cattle family in New Zealand with triplet calving ability. *Livest Sci*, 128:193-196.
- Mottershead DG, Ritter LJ, Gilchrist RB.** 2012. Signalling pathways mediating specific synergistic interactions between GDF9 and BMP15. *Mol Hum Reprod*, 18:121-128.



- Mullen MP, Hanrahan JP. 2014. Direct evidence on the contribution of a missense mutation in GDF9 to variation in ovulation rate of Finnsheep. *PLoS ONE*, 9:e95251. doi: 10.1371/journal.pone.0095251.
- Mulsant P, Lecerf F, Fabre S, Schibler L, Monget P, Lanneluc I, Pisselet C, Riquet J, Monniaux D, Callebaut I, Cribiu E, Thimonier J, Teyssier J, Bodin L, Cognie Y, Chitour N, Elsen JM. 2001. Mutation in bone morphogenetic protein receptor-1B is associated with increased ovulation rate in Booroola Merino ewes. *Proc Natl Acad Sci USA*, 98:5104-5109.
- Murakami G, Watabe T, Takaoka K, Miyazono K, Imamura T. 2003. Cooperative inhibition of bone morphogenetic protein signaling by Smurf1 and inhibitory Smads. *Mol Biol Cell*, 14:2809-2817.
- Nicol L, Bishop SC, Pong-Wong R, Bendixen C, Holm LE, Rhind SM, McNeilly AS. 2009. Homozygosity for a single base-pair mutation in the oocyte-specific GDF9 gene results in sterility in Thoka sheep. *Reproduction*, 138:921-933.
- Niswender GD, McNatty KP, Smith P, Niswender KD, Farin CE, Sawyer HR. 1990. Numbers of steroidogenic luteal cells in Booroola Merino ewes. *J Reprod Fertil*, 90:185-190.
- Nogueira MFG, Buratini J, Price CA, Castilho ACS, Pinto MGL, Barros CM. 2007. Expression of LH receptor mRNA splice variants in bovine granulosa cells: changes with follicle size and regulation by FSH in vitro. *Mol Reprod Dev*, 74:680-686.
- Peng J, Li Q, Wigglesworth K, Rangarajan A, Kattamuri C, Peterson RT, Eppig JJ, Thompson TB, Matzuk MM. 2013. Reply to Mottershead et al.: GDF9:BMP15 heterodimers are potent regulators of ovarian functions. *Proc Nat Acad Sci*, 110:E2258. doi.org/10.1073/pnas.1304497110.
- Pierson RA, Ginther OJ. 1987. Follicular populations during the estrous cycle in heifers. I. Influence of day. *Anim Reprod Sci*, 14:165-176.
- Portela VM, Dirandeh E, Guerrero-Netro HM, Zamberlam G, Barreta MH, Goetten AF, Price CA. 2015. The role of fibroblast growth factor-18 in follicular atresia in cattle. *Biol Reprod*, 92:14. doi: 10.1095/biolreprod.114.121376.
- Rajakoski E. 1960. The ovarian follicular system in sexually mature heifers with special reference to seasonal, cyclical, and left-right variations. *Acta Endocrinol (Copenh)*, 34:S7-S68.
- Reader KL, Heath DA, Lun S, McIntosh CJ, Western AH, Littlejohn RP, McNatty KP, Juengel JL. 2011. Signalling pathways involved in the cooperative effects of ovine and murine GDF9+BMP15-stimulated thymidine uptake by rat granulosa cells. *Reproduction*, 142:123-131.
- Reader KL, Mottershead DG, Martin GA, Gilchrist RB, Heath DA, McNatty KP, Juengel JL. 2016. Signalling pathways involved in the synergistic effects of human growth differentiation factor 9 and bone morphogenetic protein 15. *Reprod Fertil Dev*, 28:491-498.
- Rico C, Médigue C, Fabre S, Jarrier P, Bontoux M, Clément F, Monniaux D. 2011. Regulation of Anti-Müllerian hormone production in the cow: a multiscale study at endocrine, ovarian, follicular, and granulosa cell levels. *Biol Reprod*, 84:560-571.
- Rivera GM, Fortune JE. 2003. Selection of the dominant follicle and insulin-like growth factor (IGF)-binding proteins: Evidence that pregnancy-associated plasma protein A contributes to proteolysis of IGF-binding protein 5 in bovine follicular fluid. *Endocrinology*, 144:437-446.
- Sartorelli ES, Carvalho LM, Bergfelt DR, Ginther OJ, Barros CM. 2005. Morphological characterization of follicle deviation in Nelore (*Bos indicus*) heifers and cows. *Theriogenology*, 63:2382-2394.
- Sartori R, Fricke PM, Ferreira JCP, Ginther OJ, Wiltbank MC. 2001. Follicular deviation and acquisition of ovulatory capacity in bovine follicles. *Biol Reprod*, 65:1403-1409.
- Sartori R, Barros CM. 2011. Reproductive cycles in *Bos indicus* cattle. *Anim Reprod Sci*, 124:244-250.
- Sartori R, Gimenes LU, Monteiro Jr PLJ, Melo LF, Baruselli PS, Bastos MR. 2016. Metabolic and endocrine differences between *Bos taurus* and *Bos indicus* females that impact the interaction of nutrition with reproduction. *Theriogenology*, 86:32-40.
- Scaramuzzi RJ, Baird DT, Campbell BK, Driancourt MA, Dupont J, Fortune JE, Gilchrist RB, Martin GB, McNatty KP, McNeilly AS, Monget P, Monniaux D, Viñoles C, Webb R. 2011. Regulation of folliculogenesis and the determination of ovulation rate in ruminants. *Reprod Fertil Dev*, 23:444-467.
- Shackell GH, Hudson NL, Heath DA, Lun S, Shaw L, Condell L, Blay LR, McNatty KP. 1993. Plasma gonadotropin concentrations and ovarian characteristics in Inverdale ewes that are heterozygous for a major gene (*FecX¹*) on the X chromosome that influences ovulation rate. *Biol Reprod*, 48:1150-1156.
- Silva BDM, Castro EA, Souza CJH, Paiva SR, Sartori R, Franco MM, Azevedo HC, Silva T, Vieira AMC, Neves JP, Melo EO. 2011. A new polymorphism in the growth and differentiation factor 9 (GDF9) gene is associated with increased ovulation rate and prolificacy in homozygous sheep. *Anim Genet*, 42:89-92.
- Simões RAL, Satrapa RA, Rosa FS, Piagentini M, Castilho ACS, Ereno RL, Trinca LA, Nogueira MFG, Buratini J, Jr, Barros CM. 2012. Ovulation rate and its relationship with follicle diameter and gene expression of the LH receptor (LHR) in Nelore cows. *Theriogenology*, 77:139-147.
- Sirois J, Fortune JE. 1988. Ovarian follicular dynamics during the estrous cycle in heifers monitored by real-time ultrasonography. *Biol Reprod*, 39:308-317.
- Souza CJ, Campbell BK, Webb R, Baird DT. 1997. Secretion of inhibin A and follicular dynamics throughout the estrous cycle in the sheep with and without the Booroola gene (*FecB*). *Endocrinology*, 138:5333-5340.
- Souza CJ, MacDougall C, Campbell BK, McNeilly AS, Baird DT. 2001. The Booroola (*FecB*) phenotype is associated with a mutation in the bone morphogenetic receptor type 1 B (*BMPRI1B*) gene. *J Endocrinol*, 169:R1-6.
- Souza CJH, McNeilly AS, Benavides MV, Melo EO,



- Moraes JCF.** 2014. Mutation in the protease cleavage site of GDF9 increases ovulation rate and litter size in heterozygous ewes and causes infertility in homozygous ewes. *Anim Genet*, 45:732-739.
- Takedomi T, Kaneko H, Aoyagi Y, Konishi M, Kishi H, Watanabe G, Taya K.** 1997. Effects of passive immunization against inhibin on ovulation rate and embryo recovery in Holstein heifers. *Theriogenology*, 47:1507-1518.
- ten Dijke P and Hill CS.** 2004. New insights into TGF-beta-Smad signalling. *Trends Biochem Sci*, 29:265-273.
- Vage DI, Husdal M, Kent MP, Klemetsdal G, Boman IA.** 2013. A missense mutation in growth differentiation factor 9 (GDF9) is strongly associated with litter size in sheep. *BMC Genet*, 14:1. doi: 10.1186/1471-2156-14-1.
- Weiss A, Attisano L.** 2013. The TGFbeta superfamily signaling pathway. *Wiley Interdiscip Rev Dev Biol*, 2:47-63.
- Weller JI, Golik M, Seroussi E, Ron M, Ezra E.** 2008. Detection of quantitative trait loci affecting twinning rate in Israeli Holsteins by the daughter design. *J Dairy Sci*, 91:2469-2474.
- Wilson T, Wu XY, Juengel JL, Ross IK, Lumsden JM, Lord EA, Dodds KG, Walling GA, McEwan JC, O'Connell AR, McNatty KP, Montgomery GW.** 2001. Highly prolific Booroola sheep have a mutation in the intracellular kinase domain of bone morphogenetic protein IB receptor (ALK-6) that is expressed in both oocytes and granulosa cells. *Biol Reprod*, 64:1225-1235.
- Zhang H, Liu K.** 2015. Cellular and molecular regulation of the activation of mammalian primordial follicles: somatic cells initiate follicle activation in adulthood. *Hum Reprod Update*, 21:779-786.
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